

Regulation and Function of Fas Expression in Tumor Cells *in-vivo* and *in-vitro*

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The death receptor Fas (CD95/APO-1) appears to be involved in both pathological and normal processes of the immune system, and to play a role in tumor immunity and progression. Although Fas has been studied extensively, its precise function(s) and the regulation of its expression in tumors are not fully understood. Our lab reported that *in vitro* cultured, low Fas-expressing tumors underwent massive yet reversible up-regulation of Fas expression when injected into mice (Rosen et al., 2000). The exact regulation of Fas expression *in-vivo*, host and/or tumor factors involved are unknown. The present study was aimed at determining host factors involved in Fas up-regulation in tumor cells induced *in-vivo*, as well as examining the relationship between Fas expression and Fas activity. Fas up-regulation was accompanied by an enhanced rate of Fas-based apoptosis induced by Fas antibody, perforin-deficient CTL and by FasL trimer. Because Fas up-regulated tumor cells displayed enhanced apoptosis in response to Etoposide, increased susceptibility to apoptosis appears to be effected through the caspase pathway. Fas up-regulation was retarded when tumors were injected into immune deficient or X-irradiated mice suggesting involvement of the immune system. Compatible with this observation was the finding that Fas up-regulation could be reproduced *in-vitro* when tumor cells were co-cultured with immune spleen or peritoneal exudates cells. Fas-up regulation also occurred when low Fas expressing tumors were cultured in diluted ascitic fluids collected from tumor bearing mice. The results suggest that Fas up-regulation in tumor cells involves a soluble factors produced (e.g. nitric oxide or cytokines) and induced by the immune system. Experiments with the nitric oxide (NO) donor SNAP suggest that NO is involved in regulating Fas expression *in-vivo*.

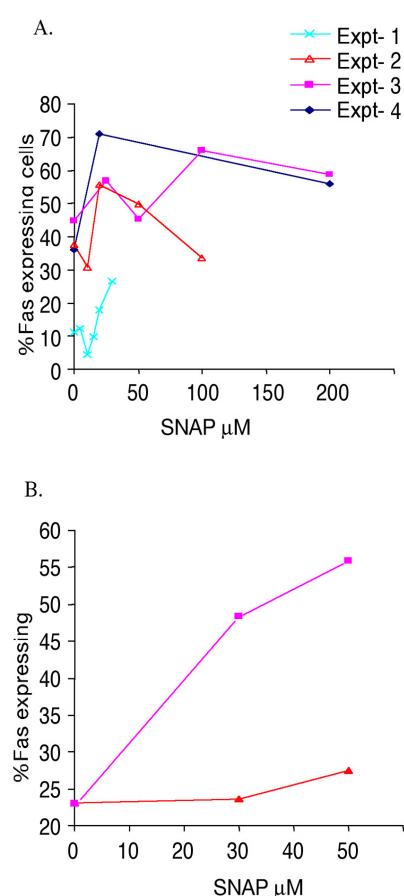


Fig. 1 The effect of the NO donor SNAP on Fas expression

A. Low Fas expressing (*cultured*) RMA-S tumor cells were exposed to 0-200 μ M of the NO donor SNAP, added once at the beginning of the experiment. After 17h incubation, cells were analyzed for Fas expression, by FACS.

B. RMA-S tumor cells were incubated in the presence of SNAP (0-50 μ M for 23h), added either once (■) at the beginning of the incubation or every 5-7h during the incubation (▲). The Fig represents one of 5 experiments.

Selected Publications

Less, H., Shilkut, M., Rubinstein, I., Berke, G. and Binah, O. (1999). Cardiac dysfunction in murine autoimmune myocarditis. *J. Autoimmunity*, 12: 209-220.

Deutsch, M., Zurgil, N., Kaufman, M. and Berke, G. (2000). Fluorescence polarization as an early measure of T Cell stimulation. In: *T Cell Protocols: Development and Activation* (Kearse, K.P. ed.), Human Press, Totowa, NJ. *Methods Mol. Biol.* 134: 221-242

Rosen, D., Li, J.H., Keidar, S., Markon, I., Orda, R. and Berke, G. (2000). Tumor immunity in perforin-deficient mice: A Role for CD95(Fas/APO-1). *J. Immunol.*, 164:3229-3235.

Shilkut, M., Gealekman, O., Rosen, D., Berke, G., Woodcock, E. and Binah, O. (2001). Electrophysiological perturbations and arrhythmogenic activity caused by activation of the Fas receptor in murine ventricular myocytes: The role of the inositol triphosphate pathway. *J. Cardiovascular Electrophysiology*, 12:185-195

Li, J-H., Rosen, D., Sondel, P. and Berke, G. (2002). Immune privilege and FasL: Two ways to inactivate effector CTLs by FasL expressing cells. *Immunology*. 105 :267-277

Yaniv, G., Shilkut, M., Lotan, R., Berke, G., Larisch, S., Binah, O. (2002). Hypoxia predisposes neonatal rat ventricular myocytes to apoptosis induced by activation of the Fas (CD95/Apo-1) receptor. *Cardiovascular Res.* 54: 611-623

Schiffenbauer, Y. S., Trubniykov, E., Zacharia, B-T., Gerbat, S., Rehavi, Z., Berke, G. and Chaitchik, S. (2002) Tumor sensitivity to anti-cancer drugs predicted by changes in fluorescence intensity and polarization *in vitro*. *Anticancer Research* 22:2663-2669

Berke, G., Krutovskikh, V. and Yamasaki H. (2003). Is mutated connexin 37 the origin of the MUT 1 and 2 octapeptides reported to be shared tumor-associated antigens of lung carcinomas 3LL and CMT? *Cancer Letters* 195: 67-72

Cohen, C. J., Denkberg, G., Segal, D., Schiffenbauer, Y., Trubniyakov, E., Berke, G. and Reiter, Y. (2003). Activation of Tumor-specific T cells by peptide-MHC complexes demonstrated at the single cell level. *J. of Immunol. Methods* 277 :39-52

Woolf, E. et al (2003) Runx 1 and 2 are required for CD8 T cell development during thymopoiesis P.N.A.S 100 :7731-7736

Berke, G. and Clark, W 2004. The killer lymphocyte Kluwer Press

Berke, G. and Clark, W 2004 CTL: Mechanism of action and role in allograft rejection In : *Immunobiology of organ transplantation* Wilkes D, and W. Burlingham Eds

Pesheh-Yaloz, N., Rosen D., Sondel P., Krammer P., and Gideon Berke. Regulation of Fas expression in tumor cell *in-vivo* and *in-vitro* (submitted)

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