

Induction in myeloid leukemic cells of genes that are expressed in different normal tissues

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Using DNA microarray and cluster analysis of expressed genes in a cloned line (M1-t-p53) of myeloid leukemic cells, we have analyzed the expression of genes that are preferentially expressed in different normal tissues. Clustering of 547 highly expressed genes in these leukemic cells showed 38 genes preferentially expressed in normal hematopoietic tissues and 122 other genes preferentially expressed in different normal nonhematopoietic tissues, including neuronal tissues, muscle, liver, and testis. We have also analyzed the genes whose expression in the leukemic cells changed after activation of WT p53 and treatment with the cytokine IL-6 or the calcium mobilizer thapsigargin. Of 620 such genes in the leukemic cells that were differentially expressed in normal tissues, clustering showed 80 genes that were preferentially expressed in hematopoietic tissues and 132 genes in different normal nonhematopoietic tissues that also included neuronal tissues, muscle, liver, and testis. Activation of p53 and treatment with IL-6 or thapsigargin induced different changes in the genes preferentially expressed in these normal tissues. These myeloid leukemic cells thus express genes that are expressed in normal nonhematopoietic tissues, and various treatments can reprogram these cells to induce other such nonhematopoietic genes. The results indicate that these leukemic cells share with normal hematopoietic stem cells the plasticity of differentiation to different cell types. It is suggested that this reprogramming to induce in malignant cells genes that are expressed in different normal tissues may be of clinical value in therapy.

Normal hematopoietic stem cells express not only hematopoietic tissue-specific genes but also genes expressed in various other normal tissues (1–3). In addition to the ability for self-renewal, normal hematopoietic stem cells thus have transcription accessibility for genes expressed in nonhematopoietic tissues, and this result can explain the plasticity of these normal stem cells (4–6). Studies on the chromosomes (7–10) and other properties (11, 12) of cancer cells have shown that cancers also have self-renewing stem cells (7–12). It was, therefore, of interest to determine whether cancer stem cells have the ability to express genes that are expressed in different normal tissues and how expression of such genes can be regulated under different conditions. To address this question, we have used a cloned line of mouse myeloid leukemic cells (M1-t-p53) (4, 13, 14) that contains a temperature-sensitive mutant p53 (V135A) protein, which changes from a mutant form at 37°C to a WT form at 32°C (15). Like the parental M1 myeloid leukemia cells (16–19), M1-t-p53 cells can be induced to undergo myeloid cell differentiation by the cytokine IL-6. When transferred from 37°C to 32°C, activation of WT p53 in M1-t-p53 cells induces apoptosis that can be effectively inhibited by IL-6 and the calcium mobilizer thapsigargin (TG) (13, 14, 18, 20).

We have previously used these cells for microarray analysis of changes in gene expression after activation of p53 in the absence or presence of IL-6 or TG (20). This analysis showed that p53-induced apoptosis can be inhibited without affecting expression of p53-regulated genes. IL-6 and TG inhibited p53-induced apoptosis by different pathways of gene expression, and IL-6 and TG induced expression of different hematopoietic differentiation-associated genes (20). DNA microarray analysis of gene

expression in a panel of 45 normal mouse tissues has shown that ≈80% of the genes are differentially expressed and that there are specific gene expression profiles in different normal tissues (21). We have now compared the gene expression in M1-t-p53 cells with the profile of gene expression in this panel of 45 normal mouse tissues. Our results indicate that these myeloid leukemic cells express genes that are preferentially expressed in different normal nonhematopoietic tissues and that activation of p53 and treatment with IL-6 or TG can induce expression of other such nonhematopoietic genes. These leukemic cells thus share with normal hematopoietic stem cells the capacity to express genes that are expressed in different normal nonhematopoietic tissues.

Materials and Methods

Cells. We have used M1-t-p53 mouse myeloid leukemic cells that express a temperature-sensitive p53 protein that changes from a mutant to a WT form after transfer from 37°C to 32°C (15) and induces apoptosis at 32°C but not at 37°C (4, 13, 14, 18, 20). The cells were cultured and treated with IL-6 or TG as described in ref. 20. Growth curves at 37°C and the formation of colonies from single cells show that M1-t-p53 and the parental M1 leukemic cells can self-renew (22).

Analysis of Tissue-Specific Gene Expression Profiles. Two DNA microarray data sets were used. In the first data set, mRNA expression levels of genes in M1-t-p53 mouse myeloid leukemic cells (20) were measured by using the Affymetrix (Santa Clara, CA) U74Av2 array, including 12,488 probe sets. In the second data set, mRNA expression profiles of genes in 45 normal mouse tissues (21) were examined by using the Affymetrix U74A array, including 12,588 probe sets. There were 9,977 probe sets in common between the two microarrays. From this point on, we refer to genes instead of probe sets. For the gene groups we discuss, the difference between the number of genes and probe sets does not exceed a few percent. For both data sets, the expression value for each gene was determined by using MICROARRAY 5.0 software (23) with default parameters. Gene expression values <20 were adjusted to 20 to eliminate noise from the data, and all values were then log₁₀-transformed.

To determine the normal expression profiles of genes that are expressed in untreated M1-t-p53 myeloid leukemic cells, we selected the most highly expressed genes in these cells, the 85th percentile, showing intensity values >314 Affymetrix fluorescence units. We then checked the expression profiles of these genes in normal mouse tissues (21). The selected genes were filtered according to their expression in the 45 normal mouse tissues to remove those genes that show a similar expression level in all 45 tissues. We used two criteria to filter the genes: (i) genes whose expression showed a high SD in the different mouse tissues, ≥0.3 of their log₁₀-transformed expression values, and (ii) genes whose expression level range was ≥1 in log₁₀-transformed values and whose expression in at least one tissue

Abbreviations: SPC, super paramagnetic clustering; TG, thapsigargin.

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Table 1. Examples of up-regulation in leukemic cells by p53, IL-6, or TG of genes that are expressed in different normal tissues

Tissues	p53				IL-6				TG			
	Gene	GenBank accession no.	Fold	<i>n</i>	Gene	GenBank accession no.	Fold	<i>n</i>	Gene	GenBank accession no.	Fold	<i>n</i>
Hematopoietic	<i>Gtse1</i>	AJ222580	11.5	22	<i>Il4ra</i>	M27960	15.0	13	<i>Adcy</i>	U12919	3.3	8
	<i>Tap1</i>	U60020	5.7		<i>Cd53</i>	X97227	4.9		<i>Hcph</i>	M68902	2.8	
Neuronal	<i>Ptpro</i>	U37465	6.2	26	<i>Snx10</i>	AI746846	3.4	1	<i>Egr2</i>	M24377	9.6	8
	<i>Pmm1</i>	AF007267	5.9						<i>Serpini1</i>	AJ001700	7.8	
Muscle	<i>Ak1</i>	AJ010108	35.6	11	<i>Sh3glb1</i>	AW125190	3.8	1	<i>Kpna1</i>	U20619	2.7	2
	<i>Lpin1</i>	AI846934	12.6						<i>Mlf1</i>	AF100171	2.0	
Liver	<i>Ccsd</i>	AI839702	4.2	8	<i>Sc5d</i>	AB016248	2.7	3	<i>Lifr</i>	D17444	4.4	3
	<i>Lyp1a</i>	AA840463	3.3		<i>Lifr</i>	D17444	2.5		<i>Cp</i>	U49430	3.3	
Testis	<i>Tcte3</i>	U21673	12.0	9	<i>Taf9</i>	AI842969	2.2	3	<i>Xmr</i>	X72697	2.6	1
	<i>Ppm1b</i>	D45859	4.6		<i>Nphp1</i>	AJ243223	2.5					

The genes are examples of up-regulated genes in M1-t-p53 leukemic cells in clusters of genes that are preferentially expressed in normal hematopoietic and nonhematopoietic tissues. Fold up-regulation is from results described by Lotem *et al.* in ref. 20. All of the genes regulated by p53, IL-6, and TG are shown in Tables 3 and 4.

50% of the cells formed colonies from single cells when cloned in agar (22). The M1-t-p53 leukemia thus has a high frequency of self-renewing stem cells.

Analysis of gene expression in mouse and human normal tissues has shown that ≈80% of the genes show a differential expression in different tissues, which can be used to identify tissue-specific gene expression profiles (21). We searched for genes that are initially highly expressed in M1-t-p53 leukemic cells and are preferentially expressed in normal nonhematopoietic tissues. We found 122 such genes in clusters whose expression profile showed preferential expression in different normal nonhematopoietic tissues, including neuronal tissues, muscle, liver, and testis. These results indicate that like normal hematopoietic stem cells, these leukemic cells express genes that are expressed in nonhematopoietic normal tissues.

The use of M1-t-p53 myeloid leukemic cells enabled us to determine the regulation of expression of nonhematopoietic genes under different conditions, including after activation of WT p53 and treatment with IL-6 or TG. We found 620 genes whose expression in the leukemic cells changed under these conditions and that were differentially expressed in different normal tissues. Of these 620 genes, 132 genes were preferentially expressed in different normal nonhematopoietic tissues that also included neuronal tissues, muscle, liver, and testis. Of these 132 genes, 59 genes were up-regulated in the leukemic cells by p53, 9 were up-regulated by IL-6, and 14 were up-regulated by TG. Only 9 of these 132 nonhematopoietic tissue-specific genes, whose expression was regulated by p53, IL-6, or TG, were among the 122 nonhematopoietic tissue-specific genes initially expressed in M1-t-p53 cells. Our results indicate that in addition to the 122 genes that are preferentially expressed in nonhematopoietic tissues and were initially expressed in the leukemic cells, p53, IL-6, and TG could reprogram these cells to change expression of other genes that are preferentially expressed in normal nonhematopoietic tissues.

The major mechanism for the transcription accessibility in the leukemic cells for genes normally expressed in nonhematopoietic tissues presumably involves changes in DNA methylation and

other epigenetic changes (19, 28–32). Induction of DNA demethylation and inhibition of histone deacetylase activity by 5-azadeoxycytidine and trichostatin A, respectively, can up-regulate expression of many genes that are silenced in cancer cells (33). It will be interesting to determine whether genes that are up-regulated by such compounds in cancer cells also include genes that are preferentially expressed in normal tissues other than the tissue in which the cancer originated. Alternative splicing may also play a role in expression of nonhematopoietic genes in the leukemic cells. One of the genes that is preferentially expressed in a nonhematopoietic tissue and is up-regulated by p53 in the M1-t-p53 leukemic cells (20) is the gene *Ak1* (adenylate kinase 1). By using the same temperature-sensitive p53 (V135A) that we used, it was shown that an alternatively spliced *Ak1* transcript can be activated by WT p53 (34). The *Mells* and *Hox11* genes that are highly expressed in some leukemias but not in normal blood cells are also alternatively spliced transcripts (35, 36). Alternatively spliced transcripts of apparently tissue-specific genes may thus be expressed in cells of another lineage.

Our results indicate that the M1 leukemic cells share with normal hematopoietic stem cells the capacity to express genes that are preferentially expressed in nonhematopoietic normal tissues. These results and the finding that under different conditions other nonhematopoietic genes were induced in these leukemic cells indicate that the leukemic cells share with normal hematopoietic stem cells the plasticity for differentiation to different cell types. Cancer therapy includes the induction of differentiation in cancer cells (16, 17, 19, 37). It will be of interest to determine to what extent different types of cancer have differences in the plasticity of gene expression for different normal tissues and how these differences will affect the growth and metastasis of the cancer and its clinical response to therapy.

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