

Microbiome at the Frontier of Personalized Medicine



Purna C. Kashyap, MBBS; Nicholas Chia, PhD; Heidi Nelson, MD; Eran Segal, PhD; and Eran Elinav, MD, PhD

CME Activity

Target Audience: The target audience for Mayo Clinic Proceedings is primarily internal medicine physicians and other clinicians who wish to advance their current knowledge of clinical medicine and who wish to stay abreast of advances in medical research.

Statement of Need: General internists and primary care physicians must maintain an extensive knowledge base on a wide variety of topics covering all body systems as well as common and uncommon disorders. Mayo Clinic Proceedings aims to leverage the expertise of its authors to help physicians understand best practices in diagnosis and management of conditions encountered in the clinical setting.

Accreditation: In support of improving patient care, Mayo Clinic College of Medicine and Science is accredited by the Accreditation Council for Continuing Medical Education (ACCME), the Accreditation Council for Pharmacy Education (ACPE), and the American Nurses Credentialing Center (ANCC) to provide continuing education for the healthcare team.

Credit Statement: Mayo Clinic College of Medicine and Science designates this journal-based CME activity for a maximum of 1.0 AMA PRA Category 1 Credit(s).™ Physicians should claim only the credit commensurate with the extent of their participation in the activity.

MOC Credit Statement: Successful completion of this CME activity, which includes participation in the evaluation component, enables the participant to earn up to 1 MOC point in the American Board of Internal Medicine's (ABIM) Maintenance of Certification (MOC) program. Participants will earn MOC points equivalent to the amount of CME credits claimed for the activity. It is the CME activity provider's responsibility to submit participant completion information to ACCME for the purpose of granting ABIM MOC credit.

Learning Objectives: On completion of this article, you should be able to (1) give examples of the role of the microbiome in drug efficacy as well as adverse events; (2) critique new studies on treatment efficacy in light of new knowledge gained from the role of the microbiome; and (3) apply knowledge of the microbiome when explaining therapeutic interventions to patients.

Disclosures: As a provider accredited by ACCME, Mayo Clinic College of Medicine and Science (Mayo School of Continuous Professional Development) must ensure balance, independence, objectivity, and scientific rigor in its educational activities. Course Director(s), Planning Committee members, Faculty, and all others who are in a position to control the content of this educational activity are required to disclose all relevant financial relationships with any commercial interest related to the subject matter of the educational activity. Safeguards against commercial bias have been put in place. Faculty also will disclose any off-label and/or investigational use of pharmaceuticals or instruments discussed in their presentation. Disclosure of this information will be published in course materials so that those participants in the activity may formulate their own judgments regarding the presentation. In their editorial and administrative roles, Karl A. Nath, MBChB, Terry L. Jopke, Kimberly D. Sankey, and Nicki M. Smith, MPA, have control of the content of this program but have no relevant financial relationship(s) with industry.

Drs Elinav and Segal are consultants to Day Two.

Method of Participation: In order to claim credit, participants must complete the following:

1. Read the activity.
2. Complete the online CME Test and Evaluation. Participants must achieve a score of 80% on the CME Test. One retake is allowed. Visit www.mayoclinicproceedings.org, select CME, and then select CME articles to locate this article online to access the online process. On successful completion of the online test and evaluation, you can instantly download and print your certificate of credit.

Estimated Time: The estimated time to complete each article is approximately 1 hour.

Hardware/Software: PC or MAC with Internet access.

Date of Release: 12/1/2017

Expiration Date: 11/30/2019 (Credit can no longer be offered after it has passed the expiration date.)

Privacy Policy: <http://www.mayoclinic.org/global/privacy.html>

Questions? Contact dctcsupport@mayo.edu.



From the Enteric Neuroscience Program, Department of Gastroenterology and Hepatology (P.C.K.), and Department of Surgery (N.C., H.N.), Mayo Clinic, Rochester, MN; and Department of Computer Science (E.S.) and Department of Immunology (E.E.), Weizmann Institute of Science, Rehovot, Israel.

Abstract

The genomic revolution promises to transform our approach to treat patients by individualizing treatments, reducing adverse events, and decreasing health care costs. The early advances using this have been realized primarily by optimizing preventive and therapeutic approaches in cancer using human genome sequencing. The ability to characterize the microbiome, which includes all the microbes that reside within and upon us and all their genetic elements, using next-generation sequencing allows us to now incorporate this important contributor to human disease into developing new preventive and therapeutic strategies. In this review we highlight the importance of the microbiome in all aspects of human disease, including pathogenesis, phenotype, prognosis, and response to treatment, as well as their role as diagnostic and therapeutic biomarkers. We provide a role for next-generation sequencing in both precise microbial identification of infectious diseases and characterization of microbial communities and their function. Taken together, the microbiome is emerging as an integral part of precision medicine approach as it not only contributes to interindividual variability in all aspects of a disease but also represents a potentially modifiable factor that is amenable to targeting by therapeutics.

© 2017 Mayo Foundation for Medical Education and Research ■ Mayo Clin Proc. 2017;92(12):1855-1864

The focus of biomedical research for most of its existence has been the ability to identify and target specific disease-associated pathways, leading to therapeutic strategies targeting a pathway. This approach remains mostly naive to interindividual variability in development of disease and response to therapy especially relevant in multifactorial diseases. However, the genomic revolution has provided a window into individual-specific information and its effect on human physiology, paving the way for personalized or precision medicine.¹ Over the past decade, efforts in oncology have allowed human genomic screening to identify a spectrum of germline-encoded sequence variations, enabling individual-specific application of preventive and therapeutic strategies. In addition to personalization of treatment based on genetic contribution to disease pathogenesis, precision medicine efforts have allowed stratification of patients based on response to treatment and development of adverse events.

The advent of microbiome research has identified the microbiome as an important contributor to human health, and in this review we highlight why the microbiome is an integral component of the precision medicine initiative (Figure). The microbiome represents the complex collection of microorganisms both within and upon us, their genomes, and collective functions.² The field has benefited vastly from the genomic revolution, allowing DNA-based identification of nonculturable bacteria inhabiting various body sites. Alteration in microbial communities (often referred to as *dysbiosis*) has been shown to be associated with diseases ranging from infectious (*Clostridium difficile* infection) to inflammatory (inflammatory bowel disease [IBD] and rheumatoid arthritis) and metabolic (diabetes and obesity) diseases, suggesting an important role for them in the pathogenesis of multifactorial conditions.³ An important aspect about the microbiome is its resilience as well as its plasticity, making it more mutable than human cells. Although on first impression these appear opposing concepts, the resilience of the microbiome is evident in health, in which, in spite of temporary insults (travel, diet, antibiotics, etc), the microbiome maintains a relatively stable steady state. In contrast, it represents a malleable organ and can be modified by dietary and other directed therapies (Figure). Furthermore, the interindividual variability in composition and metabolic capacity of the

microbiome play an important role in interactions with the environment, resulting in the development of disease as well as response to treatment and development of adverse events. The microbiome has been shown to be determined in part by the host genome, but this contribution seems small when compared with the vast environmental microbiome modulation. Hence, the important role of the microbiome in human health, the interindividual variability and contribution to host function in health, and its plasticity making it a targetable factor all point toward the importance of incorporating the microbiome into precision medicine (Figure).

The current methods use a spectrum of strategies to characterize the microbiome, the simplest being the marker gene approach using variable regions within the highly conserved 16S ribosomal RNA gene. This approach, although valuable in assessing alterations in microbial community structure, fails to provide resolution at species or strain level and does not provide sufficient functional insight into the community. Complementary approaches including metagenomics (study of all genomes in an ecosystem), metatranscriptomics (characterization of gene expression from all microbes in an ecosystem), metabolomics (characterization of all small molecule metabolites in an ecosystem), and metaproteomics (characterization of all proteins in an ecosystem) provide greater insight into functional potential as well as the expression of microbiome-derived bioactive molecules necessary to understand the therapeutic implications for the microbiome. Although the microbiome represents an attractive target for the development of personalized treatment approaches, standardization of methods to develop reliable and reproducible microbiome-based diagnostic and therapeutic strategies remains a challenge. The strong effort by the scientific community, as well as collaboration with rapidly emerging biotech companies, provides an optimistic outlook for developing microbiome-dependent and microbiome-targeted diagnostics and therapeutics.

SEQUENCING REVOLUTION ALLOWS DEVELOPMENT OF PRECISE MICROBIAL DIAGNOSTICS

Awareness of the role of the microbiome in health has both benefited from and been spurred by sequencing technology. Once considered milestone achievements requiring

the resources of dedicated genomic centers, the sequencing of a complete bacterial genome can now be performed on a laboratory bench for about a hundred dollars per sample. Rapidly declining costs and continuing development of software and algorithms for assembling genomes, either from existing reference databases or *de novo*, promise to fundamentally alter the clinical paradigm by improving our ability to track, understand, and identify disease-causing agents.⁴

Here we will describe some of the applications of bacterial genome sequencing and attempt to summarize some of the many efforts going on worldwide to bring genomic data to various problems ranging from bacterial typing^{5,6} to antiterrorism.⁷ Although these might seem like disparate use cases, what unites them is the data contained within the genome, which contains sequence variations that reflect evolutionary relationships⁸ and genes that underlie important phenotypes such as antibiotic resistance.⁹⁻¹¹

Infectious disease tracking involves the ability to detect and trace outbreaks of disease. This assists hospitals in preventing the spread of nosocomial infections, food distributors in tracing back contaminated food sources, and governments in protecting people from biological agents. The most common of these uses is in the hospital in which an indication of nosocomial disease spread can be used to improve the practice of medicine. However, these efforts have largely relied on event count and statistics; that is, they are more reactive than proactive.

The most prominent bacterial typing technique is pulse-field gel electrophoresis (PFGE),¹² which relies on restriction enzymes and gels to obtain a rough distribution of genome fragment sizes and in essence provides little detailed information and must generally be used with care and attention to detail.¹³ Great effort has been made through the standardization of PFGE techniques to enhance the comparability of results between different gels run at different laboratories.¹⁴⁻¹⁶ However, PFGE retains its difficulties in detecting infectious outbreaks across multiple centers.¹⁷

Where PFGE falls short on finer resolution and reproducibility, genome sequencing excels. Genomic data provide a base-by-base genomic “fingerprint” that enhances the resolution with which monitoring becomes possible. The fact that this may one day enable us to identify

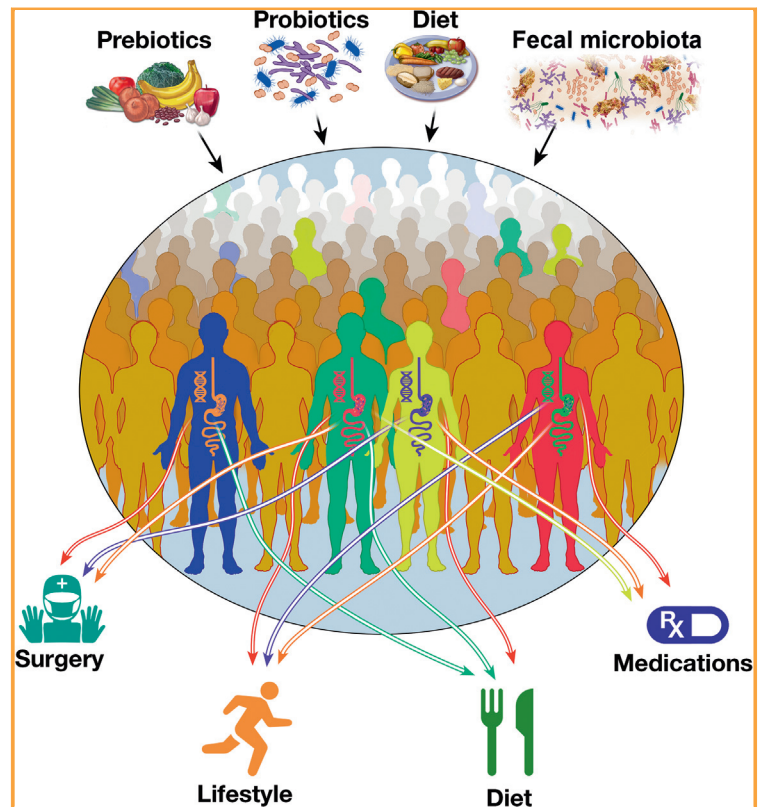


FIGURE. Gut microbiome as a determinant of human health and response to therapeutic intervention. The gut microbiome plays an important role in an individual's response to interventions ranging from dietary and lifestyle changes to medications and surgical interventions; hence, in addition to host genetics, it is important to consider the role of the gut microbiome in selecting appropriate therapy. The gut microbiome, unlike host genes, represents a modifiable factor that can be targeted by probiotics, prebiotics, diet, as well as community replacement approaches such as fecal microbiota transplant.

potential outbreaks sooner and act to prevent them before they become larger has prompted numerous studies on the efficacy of different comparison methodologies.¹⁸⁻²⁰ These methods have been tested across various species and range from single nucleotide polymorphism–based tests to use of whole-genome comparisons.^{21,22} This has also given rise to a large number of publically available phylogenetic reconstruction algorithms that analyze genomic data for signatures of relatedness to track relationships between different pathogens.²³⁻²⁵ These algorithms use the evolutionary principle of descent with modification to assess which strains descended from a recent common ancestor.

In addition, sequencing provides a great deal of information about the characteristics of an

infection. One can query for antibiotic resistance genes, identifying susceptibilities in antibiotic-resistant pathogens. This can be done using polymerase chain reaction amplicon sequencing^{26,27} or whole-genome sequencing.^{9,10} Although polymerase chain reaction–based approaches currently have an advantage in turnaround time and cost, whole-genome approaches provide more information and a common platform for evaluating multiple species. Both methods have the potential to directly assess antibiotic resistance without culture, a feature that becomes especially important in the case of slow-growing bacteria, such as tuberculosis, in which culture-based tests can take weeks to complete.

The utility of sequencing bacteria goes beyond pathogen or pathogen-complex evaluation. They can be used to directly assess more complex specimens revealing microbial ecosystems with multiple species present and represent a potential tool for diagnosing infections of unknown origin.²⁸⁻³⁰ Such broad searches require even greater bioinformatics and database support. This need has spurred a rapid growth in the number of publicly available resources for identifying potential infectious agents from complex microbiome data.^{31,32}

MICROBIOME SEQUENCING

The revolutionary change in our ability to understand the role of the microbiome came with the advent of next-generation sequencing that has allowed in-depth characterization of the gut microbiota using multi-omics approaches without the need to culture individual microbes, which in some instances can be quite challenging. The most popular method to characterize microbial communities is the marker gene approach using the 16S ribosomal RNA gene, which is highly conserved in bacteria with little evidence of horizontal gene transfer. However, this approach lacks species and strain level resolution, which often requires metagenomic sequencing and de novo assembly of genomes, providing better compositional as well as functional resolution of the microbiome.² Metatranscriptomics complements metagenomics by allowing identification of microbial genes that are expressed under different conditions. Metabolomics and metaproteomics help identify metabolites and proteins resulting from microbe-host cometabolism, which can serve as reliable biomarkers given that they represent

end products of metabolic interactions among the microbe and host. The combination of multi-omic technologies increases confidence in identified diagnostic and therapeutic biomarkers as well as provides testable hypotheses. To test emergent hypotheses generated as a result of these technologies and delineate mechanisms by which microbes influence the host, germ-free animal models provide a highly controllable experimental system with reduced complexity to study interactions between the host and its resident microbiota.

MICROBIOME AS A TOOL FOR PRECISION DIAGNOSIS AND PERSONALIZED TREATMENT STRATEGIES

There is an emerging role of the gut microbiome as a biomarker for disease phenotype, prognosis, and response to treatment in addition to the well-described associations of alterations in microbial community structure in different disease states. Inflammatory bowel disease is one of the best-studied conditions associated with dysbiosis, with the microbiome serving as an important marker of disease phenotype and response to treatment. Inflammatory bowel disease is heterogeneous with 3 major subtypes: ulcerative colitis, Crohn disease (CD), and indeterminate colitis, which not only differ in their presentation and location but also have different therapeutic strategies, making it important to obtain a precise diagnosis. The microbial populations are quite distinct even within CD with a decrease in *Faecalibacterium prausnitzii* and increase in *Escherichia coli* as well as antibodies against *E coli* outer membrane protein C seen in ileal CD compared with colonic CD^{33,34} as well as extraintestinal manifestations such as peripheral spondyloarthritis.³⁵ Gut microbiome signatures have also been associated with surgical outcomes in CD with an increase in *F prausnitzii* in the ileal mucosa associated with decreased disease recurrence at 6 months. In spite of several studies highlighting changes in the microbiome in IBD, there is lack of agreement among studies, making it imperative to have large cohorts from different geographic locations to overcome the effect of disease subtype, antibiotic use, diet, and other factors affecting the gut microbiome. This was highlighted in a study of treatment-naïve patients with CD, in which a large patient cohort was needed to identify discriminatory taxa.³⁶ The study further found the need to study mucosa-associated bacteria, which may be

more relevant in inflammatory diseases such as IBD. In addition to IBD, microbiome signatures have been described in several other gastrointestinal diseases. *Fusobacterium nucleatum* has been implicated in colorectal cancer through its FadA adhesion serving as both a diagnostic and a therapeutic marker.³⁷ *Clostridium difficile* infection has been associated with decreased microbial diversity and a decrease in secondary bile acid production.³⁸ In addition, recently 2 studies have identified microbiome signatures in *Clostridium difficile* infection that allow prediction of disease outcome enabling therapeutic stratification.^{39,40} An expansion of Proteobacteria in the setting of dysbiotic microbiota was described in patients with celiac disease with gastrointestinal symptoms compared with those with extraintestinal manifestations of celiac disease.⁴¹ In addition to diseases within the gastrointestinal tract, it is interesting to note that several studies have described gut microbiome signatures in systemic disorders such as rheumatoid arthritis. An expansion of *Prevotella copri* has been described in new-onset rheumatoid arthritis.⁴² Another recent study identified enrichment of *Collinsella*, *Eggerthella*, and *Faecalibacterium* in patients with rheumatoid arthritis and a strong association of *Collinsella* with high levels of α -amino adipic acid and asparagine as well as production of the α -amino adipic cytokine interleukin 17A and experimental arthritis.⁴³ These few examples are just a window into accumulating experimental evidence for the role of the microbiome in human disease and the future of microbiome-based diagnostic and therapeutic biomarkers. Although these studies are helpful in identifying biomarkers, much work still needs to be done in validating these signatures in large multicenter cohorts as well identifying potential causative role using a combination of in vitro and in vivo models.

MICROBIOME AS A DETERMINANT OF HUMAN THERAPEUTICS

The ecology of a microbial population, as in any ecosystem, involves a lot of cross talk between different species. Microbial survival and growth is governed strongly by their chemical environment, and unsurprisingly, they have evolved gene cassettes for chemical warfare.^{44,45} Indeed, the discovery of antibiotics first occurred in microbial culture as a unique characteristic of colonies⁴⁶ and since then broader surveys of the soil microbiota have revealed an even greater array

of antibiotic compounds.^{47,48} Recently, this has been extended to the human microbiome as well across multiple sites along the human body,⁴⁹ which means the source of compounds we need to harness control over our microbiome might already be within us.

In addition to antibiotics and signaling agents, the discovery of the so-called bacterial immune system, that is, the CRISPR-Cas system, allows bacteria to resist and exclude bacteriophages from the population by targeting specific sequences for cleavage.⁵⁰ Although providing adaptive immunity to viruses, the industrial uses of this biological system have been widely recognized, leading to the implementation of synthetic CRISPR-Cas systems⁵¹ that have led to the implementation of species-specific antimicrobial agents⁵² that may be able to preserve the bulk of the microbiome while still making key alterations.

In addition to being a source of therapeutics with implications for human disease, the microbiome serves as both a modulator of traditional therapies and a target for therapies. The interindividual variability in response to therapy and development of adverse events has been attributed to individual specific disease phenotype and host genetics, but gut microbiota is often overlooked. However, the gut microbiota plays an important role in drug transformation affecting their efficacy. Acetaminophen, a commonly used analgesic drug, may compete with bacteria-generated *p*-cresol for O-sulfonation, resulting in acetaminophen glucuronidation, which can explain in part interindividual variability in analgesic response⁵³ as well as differences in adverse events due to accumulation of its toxic metabolite *N*-acetyl-*p*-benzoquinone imine. Microbiome markers of drug efficacy ranging from chemotherapeutic agents to statins have been widely described. *Bifidobacterium* has been found to augment tumor control in mouse models of melanoma treated with anti-programmed death-ligand 1.⁵⁴ Similarly in humans, *Bacteroides* have been suggested to be responsible for antitumor effects of cytotoxic T-lymphocyte associated protein 4 blockade, commonly used for cancer immunotherapy.⁵⁵ Irinotecan (7-ethyl-10-[4-(1-piperidino)-1-piperidino]carbonyloxycamptothecin), a chemotherapeutic agent commonly used for colorectal cancer, can undergo β -glucuronidation by gut bacteria, resulting in an active metabolite that causes severe diarrhea.⁵⁶ It is important to

note that host genetic variation also plays an important role in shaping the gut microbiome.⁵⁷ Bacteria-derived coprostanol levels have been associated with clinical response to statins, which are commonly used as low-density lipoprotein cholesterol—lowering agents. Digoxin, a cardiac glycoside with a narrow therapeutic window, can be inactivated by *Eggerthella lenta* in the gut. Finally, a recent study highlights the role of the gut microbiota in mediating the antidiabetic effects of metformin.⁵⁸ These examples clearly highlight the importance of considering the gut microbiota when determining drug responses akin to pharmacogenomics (Figure). The combination of the 2 approaches will allow us to impart more precise and effective therapeutics while decreasing overall adverse events.

TARGETING THE MICROBIOME TO IMPROVE HEALTH

In addition to serving as diagnostic and therapeutic biomarkers and modulating therapeutic responses to drugs, the appealing aspect of the microbiome is its plasticity and our ability to modify components of the microbiome. The traditional approach to target microbial populations has been with the use of antibiotics, which are both essential and effective for treating systemic infections typically resulting from pathogen invasion. However, the unintended off-target effects on microbial community structure as well as adverse effects in humans makes it less appealing as precise therapies to target the microbiome.⁵⁹⁻⁶¹ There is a continued role for developing pathogen-targeted antibiotics by identifying specific targets, which narrow the spectrum of the antibiotic. A novel approach includes mining the microbiota for therapeutic targets by identifying specific functions that affect the host, allowing us to modify microbial community functionality without harming the community itself.⁶² An example is the role of trimethylamine oxidase in atherosclerosis and the inhibition of bacterial trimethylamine lyases by 3,3-dimethyl-1-butanol, decreasing bacterial trimethylamine production in a high choline diet—fed murine model.⁶³ Even with precise targeting of a single pathway, there were still alterations in the microbiome by 3,3-dimethyl-1-butanol, highlighting the complexity of microbial interactions within these ecosystems. There are several other approaches to target the microbiome, including use of probiotics, prebiotics, as well as dietary interventions. The early probiotics (live microorganisms that, when

given in sufficient amounts, confer a health benefit on the host) were dominated by members of the genera *Lactobacillus* and *Bifidobacterium*, but lacked precision in terms of targeting a biological function. A recent systematic review of medium- to high-quality randomized controlled trials using probiotics found that there was no significant effect on the gut microbiota compared to placebo. The clinical efficacy of currently available probiotics is difficult to assess given the small sample sizes limiting the power, heterogeneity in strains of bacteria used, end points, duration of treatment and molecular methods of studying the gut microbiota, recording of baseline measurements such as diet, and often a lack of good preclinical mechanistic data.⁶⁴ However, recent work highlights the promise of next-generation probiotics that will be developed using targeted approaches to alter microbial metabolism in a disease-specific manner. A precision approach using *Clostridium scindens* to augment resistance to *C difficile* infection by targeting secondary bile acid pathway³⁸ is one such example. Similarly, a multi-component probiotic was shown to modulate the gut microbiome with resultant suppression of hepatocellular carcinoma in a mouse model.⁶⁵ The advent of genetic engineering and synthetic biology approaches also hold promise for the development of precision probiotics.⁶⁶ An example is the engineering of a common gut commensal to secrete the molecular signal cholera autoinducer-1, inhibiting *Vibrio cholerae* virulence in a mouse model.⁶⁷ Furthermore, tunable expression tools in robust colonizers of the human gut provides us the ability to further calibrate delivery of bioactive compounds by these precision probiotics.⁶⁸ Prebiotic (ingredients that are selectively fermented by gut microbes and confer a health benefit) approaches aim to modulate the microbial community in a way that is beneficial to human health. Although the early prebiotics have focused on promoting the growth of a single or group of beneficial bacteria, they fail to account for the downstream effects on other microbial members. Similar to probiotics, prebiotics that are mainly composed of fermentable oligosaccharides such as inulin and fructooligosaccharides have focused on increasing growth of potentially beneficial bacteria such as *Bifidobacterium*. The lack of an ecosystem approach is reflected in the modest clinical efficacy of available prebiotics. The development of next-generation prebiotics will require careful modeling of the metabolic interactions among the members of the ecosystem to better understand the overall effects

on the community and host physiology. Fecal microbiota transplant (FMT) that entails transfer of the healthy gut microbiota from a donor either orally via capsules or endoscopically has been highly successful as an ecosystem approach in treating recurrent *C difficile* infection.⁶⁹ A similar approach with FMT has been tested in multiple diseases associated with microbiome alterations but has failed to show clinical efficacy. However, the use of FMT for diseases such as IBD has provided insight into donor specificity⁷⁰ in terms of response, suggesting a role of individualizing FMT approaches in multifactorial diseases such as IBD, in contrast to the approach in *C difficile* infection.

Finally, diet has major implications for the microbiome as it is the primary nutrient source of microbes. Dietary manipulations fall with 3 distinct approaches. The use of microbiome markers in optimizing dietary interventions, modulating the diet based on the microbiome and using diet to alter the microbiome. Dietary interventions limiting fermentable oligosaccharides, disaccharides, monosaccharides, and polyols have shown to be beneficial in ameliorating symptoms in patients with irritable bowel syndrome.⁷¹ However, long-term use of such an intervention can decrease microbial short chain fatty acid production, which, in turn, may have negative implications for human health. A recent study identified microbiome markers that predict a positive response to fermentable oligosaccharides, disaccharides, monosaccharides, and polyols⁷² with the potential to allow optimization of therapy and minimizing undesirable adverse effects in individuals less likely to respond. An important aspect of the gut microbiome is its role in determining host responses to dietary components given that the microbiome plays an important role in metabolism of dietary nutrients. Zeevi et al⁷³ found large interpersonal differences in postprandial glycemic responses to dietary components in an elegant study of 800 participants. The prediction engine used to make the predictions incorporated multiple host and microbial parameters, and they found that the incorporation of microbiome-derived features improved the accuracy of prediction of glycemic responses.⁷³ In a follow-up study, the authors found significant interpersonal variability in the glycemic response to different bread types, and the glycemic response to different types of bread could be predicted solely from microbiome data before

the intervention.⁷⁴ These studies highlight the ability to personalize nutritional intervention to improve host physiology based on an individual's microbiome. It is important to note that both short-term and long-term dietary patterns have a significant effect on shaping the microbiome. A diet high in protein and fat in the long-term has been associated with enrichment of *Bacteroides*, whereas a carbohydrate-rich diet has been associated with *Prevotella*.⁷⁵ Sonnenburg et al⁷⁶ reported that a Western diet low in microbiota accessible carbohydrates leads to decreased diversity in the microbiota of humanized mice, which are largely reversible within a single generation, but over several generations, this leads to a progressive loss of diversity that cannot be recovered by diet alone and needs replacement of the microbiota. This has significant implications for populations consuming a Western diet, which has been associated with decreased diversity and an increase in autoimmune diseases. The study suggests that even long-term dietary effects may be reversible within a certain time frame. Interestingly, short-term dietary effects on the microbiome seem to be easily reversible even when using extreme dietary interventions.⁷⁷ Moreover, short-term dietary interventions have shown to have beneficial effects on the host and gut microbiome. In the study by Zeevi et al⁷³ mentioned previously, introduction of meals associated with low postprandial glucose response led to an increase in bacteria thought to be protective against type 2 diabetes mellitus such as *Roseburia inulinivorans*, *Eubacterium eligens*, and *Bacteroides vulgatus*. Similarly, a 3-day dietary intervention with barley-based bread was associated with higher *Prevotella/Bacteroides* ratio and improved glucose metabolism.⁷⁸ It is interesting to note that changes in gut microbiota to a similar dietary intervention can vary depending on an individual's microbiome.⁷⁹ Taken together, it is apparent that although the relationship of diet and gut microbiome is complex, it is highly relevant in determining host responses to diet as well as predicting changes in the microbiome in response to the diet.

CONCLUSION

In this review we highlight the importance of incorporating the microbiome as a component of personalized or precision medicine to improve diagnosis, reduce disease risk, and

optimize early detection and treatment. Microbial fingerprints could serve as precise, noninvasive, accessible, and economic tools that could be used for personalized disease diagnosis including phenotypes, severity, and prognosis. The role of the microbiome in the metabolism of many chemical compounds makes it a key player in determining drug availability, efficacy, and toxicity, making it indispensable for developing personalized drug therapies. Finally, the ability to manipulate the microbiome makes it appealing in developing personalized treatment approaches by using precision microbiome targeting approaches. The use of approaches targeting specific microbial pathways tailored to an individual's microbiota may enable the development of treatment of multifactorial disorders such as IBD, obesity, and diabetes mellitus. The development of precision probiotics using genetic engineering approaches, next-generation prebiotics resulting from a better understanding of metabolic interactions among members of the microbial ecosystem, and personalized dietary therapies tailored to an individual's microbiota will form the new frontier in the field of personalized medicine.

Overall, the outlook is optimistic, but there are also substantial challenges in the field. To implement microbiome-based diagnostics and therapeutics, we need to develop uniform collection, sequencing, and analysis standards that would enhance reproducibility of results across centers and reduce biases in their interpretation. Most current studies are based on disease association, but we need to better define the mechanisms by which microbiota influence aspects of human disease to develop more reliable biomarkers. Furthermore, we are only beginning to appreciate the contribution of other microorganisms such as fungi, bacteriophages, and parasites as well as the interkingdom signaling among the microorganisms and the host. As we unravel aspects of these complex interactions, we will begin to develop more robust strategies to address the effect of the microbiome on the host.

The plasticity of the microbiome, while being advantageous in terms of making it amenable to intervention, also poses a challenge in terms of stability of changes. This was highlighted above, wherein dietary interventions can be developed on the basis of an individual's microbiome; however, it has the

potential to change the microbiome itself. Hence, a systems approach to better understand the diet-microbiome interaction will allow the identification of dependencies between dietary compounds and bacterial taxa as well as prediction of trends in their variation resulting from dietary intervention.

These challenges apart, the integration of microbiome-based diagnostics and therapeutics into other components of personalized medicine such as pharmacogenomics and epigenomics will be an integral part of the new era in patient care. This integration will further enhance our ability to find the right treatment for the right patient while, at the same time, reducing adverse events and health care cost.

ACKNOWLEDGMENTS

We thank Lyndsay Busby, AS, for her administrative assistance.

Abbreviations and Acronyms: CD = Crohn disease; FMT = fecal microbiota transplant; IBD = inflammatory bowel disease; PFGE = pulse-field gel electrophoresis

Grant Support: The work was supported in part by grants R01 DK114007 and R03 DK111850 (P.C.K.) from the National Institutes of Health.

Potential Competing Interests: Eran Elinav and Eran Segal are consultants to Day Two.

Correspondence: Address to Puma C. Kashyap, MBBS, Enteric Neuroscience Program, Division of Gastroenterology and Hepatology, Mayo Clinic, 200 First St SW, Rochester, MN 55905 (kashyap.puma@mayo.edu). Individual reprints of this article and a bound reprint of the entire Symposium on Precision Medicine will be available for purchase from our website www.mayoclinicproceedings.org.

The Symposium on Precision Medicine will continue in an upcoming issue.

REFERENCES

1. Jameson JL, Longo DL. Precision medicine—personalized, problematic, and promising. *N Engl J Med*. 2015;372(23):2229-2234.
2. Marchesi JR, Ravel J. The vocabulary of microbiome research: a proposal. *Microbiome*. 2015;3:31.
3. Marchesi JR, Adams DH, Fava F, et al. The gut microbiota and host health: a new clinical frontier. *Gut*. 2016;65(2):330-339.
4. Didelot X, Bowden R, Wilson DJ, Peto TE, Crook DW. Transforming clinical microbiology with bacterial genome sequencing. *Nat Rev Genet*. 2012;13(9):601-612.
5. Köser CU, Ellington MJ, Cartwright EJP, et al. Routine use of microbial whole genome sequencing in diagnostic and public health microbiology. *PLoS Pathog*. 2012;8(8):e1002824.
6. Joensen KG, Scheut F, Lund O, et al. Real-time whole-genome sequencing for routine typing, surveillance, and outbreak

- detection of verotoxigenic *Escherichia coli*. *J Clin Microbiol*. 2014; 52(5):1501-1510.
7. Keim P, Van Ert MN, Pearson T, Vogler AJ, Huynh LY, Wagner DM. Anthrax molecular epidemiology and forensics: using the appropriate marker for different evolutionary scales. *Infect Genet Evol*. 2004;4(3):205-213.
 8. Harris SR, Feil EJ, Holden MT, et al. Evolution of MRSA during hospital transmission and intercontinental spread. *Science*. 2010; 327(5964):469-474.
 9. Leopold SR, Goering RV, Witten A, Harmsen D, Mellmann A. Bacterial whole-genome sequencing revisited: portable, scalable, and standardized analysis for typing and detection of virulence and antibiotic resistance genes. *J Clin Microbiol*. 2014; 52(7):2365-2370.
 10. Veenemans J, Overvest IT, Snelders E, et al. Next-generation sequencing for typing and detection of resistance genes: performance of a new commercial method during an outbreak of extended-spectrum- β -lactamase-producing *Escherichia coli*. *J Clin Microbiol*. 2014;52(7):2454-2460.
 11. Machado MP, Ribeiro-Gonçalves B, Silva M, Ramirez M, Camiço JA. Epidemiological surveillance and typing methods to track antibiotic resistant strains using high throughput sequencing. *Methods Mol Biol*. 2017;1520:331-356.
 12. Schwartz DC, Cantor CR. Separation of yeast chromosome-sized DNAs by pulsed field gradient gel electrophoresis. *Cell*. 1984;37(1):67-75.
 13. Foxman B, Zhang L, Koopman JS, Manning SD, Marrs CF. Choosing an appropriate bacterial typing technique for epidemiologic studies. *Epidemiol Perspect Innov*. 2005;2:10.
 14. Swaminathan B, Barrett TJ, Hunter SB, Tauxe RV; CDC PulseNet Task Force. PulseNet: the molecular subtyping network for foodborne bacterial disease surveillance, United States. *Emerg Infect Dis*. 2001;7(3):382-389.
 15. Kam KM, Luey CK, Parsons MB, et al; Vibrio parahaemolyticus PulseNet PFGE Protocol Working Group. Evaluation and validation of a PulseNet standardized pulsed-field gel electrophoresis protocol for subtyping *Vibrio parahaemolyticus*: an international multicenter collaborative study. *J Clin Microbiol*. 2008;46(8):2766-2773.
 16. Scharff RL, Besser J, Sharp DJ, Jones TF, Peter GS, Hedberg CW. An economic evaluation of PulseNet: a network for foodborne disease surveillance. *Am J Prev Med*. 2016;50(5 Suppl 1):S66-S73.
 17. Price JR, Didelot X, Crook DW, Llewelyn MJ, Paul J. Whole genome sequencing in the prevention and control of *Staphylococcus aureus* infection. *J Hosp Infect*. 2013;83(1):14-21.
 18. Viau RA, Kiedrowski LM, Kreiswirth BN, et al. A comparison of molecular typing methods applied to *Enterobacter cloacae* complex: hsp60 sequencing, Rep-PCR, and MLST. *Pathog Immun*. 2017;2(1):23-33.
 19. Salipante SJ, SenGupta DJ, Cummings LA, Land TA, Hoogestraat DR, Cookson BT. Application of whole-genome sequencing for bacterial strain typing in molecular epidemiology. *J Clin Microbiol*. 2015;53(4):1072-1079.
 20. Jolley KA, Maiden MC. Automated extraction of typing information for bacterial pathogens from whole genome sequence data: *Neisseria meningitidis* as an exemplar. *Euro Surveill*. 2013;18(4):20379.
 21. Coll F, McNemey R, Guerra-Assunção JA, et al. A robust SNP barcode for typing *Mycobacterium tuberculosis* complex strains. *Nat Commun*. 2014;5:4812.
 22. Cunningham SA, Chia N, Jeraldo PR, et al. Comparison of whole-genome sequencing methods for analysis of three methicillin-resistant *Staphylococcus aureus* outbreaks. *J Clin Microbiol*. 2017;55(6):1946-1953.
 23. Feil EJ, Li BC, Aanensen DM, Hanage WP, Spratt BG. eBURST: inferring patterns of evolutionary descent among clusters of related bacterial genotypes from multilocus sequence typing data. *J Bacteriol*. 2004;186(5):1518-1530.
 24. Vaz C, Francisco AP, Silva M, et al. TypOn: the microbial typing ontology. *J Biomed Semantics*. 2014;5(1):43.
 25. Nascimento M, Sousa A, Ramirez M, Francisco AP, Camiço JA, Vaz C. PHYLOViZ 2.0: providing scalable data integration and visualization for multiple phylogenetic inference methods. *Bioinformatics*. 2017;33(1):128-129.
 26. Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG. Multi-locus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J Clin Microbiol*. 2000;38(3):1008-1015.
 27. Ng LK, Martin I, Alfa M, Mulvey M. Multiplex PCR for the detection of tetracycline resistant genes. *Mol Cell Probes*. 2001;15(4):209-215.
 28. Wilson MR, Naccache SN, Samayoa E, et al. Actionable diagnosis of neuroleptospirosis by next-generation sequencing. *N Engl J Med*. 2014;370(25):2408-2417.
 29. Scholz CF, Jensen A. Development of a Single Locus Sequence Typing (SLST) scheme for typing bacterial species directly from complex communities. *Methods Mol Biol*. 2017;1535:97-107.
 30. Sahi JW, Schupp JM, Rasko DA, Colman RE, Foster JT, Keim P. Phylogenetically typing bacterial strains from partial SNP genotypes observed from direct sequencing of clinical specimen metagenomic data. *Genome Med*. 2015;7(1):52.
 31. Wattam AR, Abraham D, Dalay O, et al. PATRIC, the bacterial bioinformatics database and analysis resource. *Nucleic Acids Res*. 2014;42(Database issue):D581-D591.
 32. Zolfo M, Tett A, Jousson O, Donati C, Segata N. MetaMLST: multi-locus strain-level bacterial typing from metagenomic samples. *Nucleic Acids Res*. 2017;45(2):e7.
 33. Willing B, Halfvarson J, Dicksved J, et al. Twin studies reveal specific imbalances in the mucosa-associated microbiota of patients with ileal Crohn's disease. *Inflamm Bowel Dis*. 2009;15(5):653-660.
 34. Willing BP, Dicksved J, Halfvarson J, et al. A pyrosequencing study in twins shows that gastrointestinal microbial profiles vary with inflammatory bowel disease phenotypes. *Gastroenterology*. 2010;139(6):1844-1854.e1841.
 35. Viladomiu M, Kivolowitz C, Abdulhamid A, et al. IgA-coated *E. coli* enriched in Crohn's disease spondyloarthritis promote TH17-dependent inflammation. *Sci Transl Med*. 2017;9(376).
 36. Gevers D, Kugathasan S, Denson LA, et al. The treatment-naïve microbiome in new-onset Crohn's disease. *Cell Host Microbe*. 2014;15(3):382-392.
 37. Rubinstein MR, Wang X, Liu W, Hao Y, Cai G, Han YW. Fusobacterium nucleatum promotes colorectal carcinogenesis by modulating E-cadherin/ β -catenin signaling via its FadA adhesin. *Cell Host Microbe*. 2013;14(2):195-206.
 38. Buffie CG, Bucci V, Stein RR, et al. Precision microbiome reconstitution restores bile acid mediated resistance to *Clostridium difficile*. *Nature*. 2015;517(7533):205-208.
 39. Khanna S, Montassier E, Schmidt B, et al. Gut microbiome predictors of treatment response and recurrence in primary *Clostridium difficile* infection. *Aliment Pharmacol Ther*. 2016;44(7):715-727.
 40. Seekatz AM, Rao K, Santhosh K, Young VB. Dynamics of the fecal microbiome in patients with recurrent and nonrecurrent *Clostridium difficile* infection. *Genome Med*. 2016;8(1):47.
 41. Wacklin P, Kaukinen K, Tuovinen E, et al. The duodenal microbiota composition of adult celiac disease patients is associated with the clinical manifestation of the disease. *Inflamm Bowel Dis*. 2013;19(5):934-941.
 42. Scher JU, Sczesnak A, Longman RS, et al. Expansion of intestinal *Prevotella copri* correlates with enhanced susceptibility to arthritis. *Elife*. 2013;2:e01202.
 43. Chen J, Wright K, Davis JM, et al. An expansion of rare lineage intestinal microbes characterizes rheumatoid arthritis. *Genome Med*. 2016;8(1):43.
 44. Shank EA, Kolter R. New developments in microbial interspecies signaling. *Curr Opin Microbiol*. 2009;12(2):205-214.
 45. Cornforth DM, Foster KR. Antibiotics and the art of bacterial war. *Proc Natl Acad Sci U S A*. 2015;112(35):10827-10828.
 46. Jones D, Metzger HJ, Schatz A, Waksman SA. Control of gram-negative bacteria in experimental animals by streptomycin. *Science*. 1944;100(2588):103-105.

47. Aigle B, Lautre S, Spittler D, et al. Genome mining of *Streptomyces ambofaciens*. *J Ind Microbiol Biotechnol*. 2014;41(2):251-263.
48. Bachmann BO, Van Lanen SG, Baltz RH. Microbial genome mining for accelerated natural products discovery: is a renaissance in the making? *J Ind Microbiol Biotechnol*. 2014;41(2):175-184.
49. Donia MS, Cimemancic P, Schulze CJ, et al. A systematic analysis of biosynthetic gene clusters in the human microbiome reveals a common family of antibiotics. *Cell*. 2014;158(6):1402-1414.
50. Barrangou R, Fremaux C, Deveau H, et al. CRISPR provides acquired resistance against viruses in prokaryotes. *Science*. 2007;315(5819):1709-1712.
51. Cong L, Ran FA, Cox D, et al. Multiplex genome engineering using CRISPR/Cas systems. *Science*. 2013;339(6121):819-823.
52. Bikard D, Euler CW, Jiang W, et al. Exploiting CRISPR-Cas nucleases to produce sequence-specific antimicrobials. *Nat Biotechnol*. 2014;32(11):1146-1150.
53. Clayton TA, Baker D, Lindon JC, Everett JR, Nicholson JK. Pharmacometabonomic identification of a significant host-microbiome metabolic interaction affecting human drug metabolism. *Proc Natl Acad Sci U S A*. 2009;106(34):14728-14733.
54. Sivan A, Corrales L, Hubert N, et al. Commensal *Bifidobacterium* promotes antitumor immunity and facilitates anti-PD-L1 efficacy. *Science*. 2015;350(6264):1084-1089.
55. Vétizou M, Pitt JM, Daillère R, et al. Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. *Science*. 2015;350(6264):1079-1084.
56. Wallace BD, Wang H, Lane KT, et al. Alleviating cancer drug toxicity by inhibiting a bacterial enzyme. *Science*. 2010;330(6005):831-835.
57. Blekhan R, Goodrich JK, Huang K, et al. Host genetic variation impacts microbiome composition across human body sites. *Genome Biol*. 2015;16:191.
58. Wu H, Esteve E, Tremaroli V, et al. Metformin alters the gut microbiome of individuals with treatment-naïve type 2 diabetes, contributing to the therapeutic effects of the drug. *Nat Med*. 2017;23(7):850-858.
59. Rubinstein E, Camm J. Cardiotoxicity of fluoroquinolones. *J Antimicrob Chemother*. 2002;49(4):593-596.
60. Galatti L, Giustini SE, Sessa A, et al. Neuropsychiatric reactions to drugs: an analysis of spontaneous reports from general practitioners in Italy. *Pharmacol Res*. 2005;51(3):211-216.
61. Rubin BK, Tamaoki J. *Antibiotics as Anti-Inflammatory and Immunomodulatory Agents*. Boston: Birkhäuser; 2005.
62. Wallace BD, Redinbo MR. The human microbiome is a source of therapeutic drug targets. *Curr Opin Chem Biol*. 2013;17(3):379-384.
63. Wang Z, Roberts AB, Buffa JA, et al. Non-lethal inhibition of gut microbial trimethylamine production for the treatment of atherosclerosis. *Cell*. 2015;163(7):1585-1595.
64. Kristensen NB, Bryrup T, Allin KH, Nielsen T, Hansen TH, Pedersen O. Alterations in fecal microbiota composition by probiotic supplementation in healthy adults: a systematic review of randomized controlled trials. *Genome Med*. 2016;8(1):52.
65. Li J, Sung CY, Lee N, et al. Probiotics modulated gut microbiota suppresses hepatocellular carcinoma growth in mice. *Proc Natl Acad Sci U S A*. 2016;113(9):E1306-E1315.
66. Amalaradjou MA, Bhunia AK. Bioengineered probiotics, a strategic approach to control enteric infections. *Bioengineered*. 2013;4(6):379-387.
67. Ruder WC, Lu T, Collins JJ. Synthetic biology moving into the clinic. *Science*. 2011;333(6047):1248-1252.
68. Whitaker WR, Shepherd ES, Sonnenburg JL. Tunable Expression Tools Enable Single-Cell Strain Distinction in the Gut Microbiome. *Cell*. 2017;169(3):538-546.e512.
69. Kelly CR, Kahn S, Kashyap P, et al. Update on fecal microbiota transplantation 2015: indications, methodologies, mechanisms, and outlook. *Gastroenterology*. 2015;149(1):223-237.
70. Moayyedi P, Surette MG, Kim PT, et al. Fecal microbiota transplantation induces remission in patients with active ulcerative colitis in a randomized controlled trial. *Gastroenterology*. 2015;149(1):102-109.e106.
71. Halmos EP, Power VA, Shepherd SJ, Gibson PR, Muir JG. A diet low in FODMAPs reduces symptoms of irritable bowel syndrome. *Gastroenterology*. 2014;146(1):67-75.e65.
72. Bennet SM, Böhn L, Störsrud S, et al. Multivariate modelling of faecal bacterial profiles of patients with IBS predicts responsiveness to a diet low in FODMAPs [published online ahead of print April 17, 2017]. *Gut*. <https://doi.org/10.1136/gutjnl-2016-313128>.
73. Zeevi D, Korem T, Zmora N, et al. Personalized nutrition by prediction of glycemic responses. *Cell*. 2015;163(5):1079-1094.
74. Korem T, Zeevi D, Zmora N, et al. Bread affects clinical parameters and induces gut microbiome-associated personal glycemic responses. *Cell Metab*. 2017;25(6):1243-1253.e1245.
75. Wu GD, Chen J, Hoffmann C, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science*. 2011;334(6052):105-108.
76. Sonnenburg ED, Smits SA, Tikhonov M, Higginbottom SK, Wingreen NS, Sonnenburg JL. Diet-induced extinctions in the gut microbiota compound over generations. *Nature*. 2016;529(7585):212-215.
77. David LA, Maurice CF, Carmody RN, et al. Diet rapidly and reproducibly alters the human gut microbiome. *Nature*. 2014;505(7484):559-563.
78. Kovatcheva-Datchary P, Nilsson A, Akrami R, et al. Dietary fiber-induced improvement in glucose metabolism is associated with increased abundance of *Prevotella*. *Cell Metab*. 2015;22(6):971-982.
79. Smits SA, Marcobal A, Higginbottom S, Sonnenburg JL, Kashyap PC. Individualized responses of gut microbiota to dietary intervention modeled in humanized mice. *mSystems*. 2016;1(5).