

Type I Diabetes Mellitus, Infection and Toll-Like Receptors

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1. EPIDEMIOLOGY OF TYPE-I DIABETES

During the last forty years, the incidence of type I diabetes mellitus (T1DM) has shown a significant increase in developed countries [1]. Gross geographical differences can be seen in the increase of T1DM, and Europe provides us with a good example [1]. Northern European countries have higher incidences of T1DM than southern European countries, with Finland showing the highest European (and world) incidence [2]. The genetic background is an important factor in determining the susceptibility to autoimmune disease [3], however several observations indicate that genetic differences cannot account for the uneven distribution of T1DM. First, epidemiological studies carried out in Northern Ireland [4] and England [5] have found a positive correlation between a low incidence of type I diabetes and poorer socioeconomic conditions. Second, populations migrating between countries with different values of T1DM incidence are interesting examples: children born to parents who migrated to Yorkshire from Pakistan show an incidence of T1DM indistinguishable from that seen among non-migrants in England (11.7/100,000), which is significantly higher than Pakistan's incidence of T1DM (1/100,000) [1, 6, 7]. Third, in a recent paper Hyttinen and co-workers report that after studying 22,650 twins, the concordance rate for T1DM was only 27.3% in monozygotic twins and 3.8% in dizygotic twins [2]. In summary, these data highlight the importance of environmental factors in the worldwide increase of T1DM.

2. THE HYGIENE HYPOTHESIS

The hygiene hypothesis associates the increase in the incidence of autoimmune diseases and allergy in developed countries with the effects of the environment on the immune system. It postulates that childhood infections educate the immune system on how to react to antigenic challenge. The immune system is a self-organizing system and like the brain it requires experience to learn how to behave [8]. However, as a consequence of improved hygiene, vaccination campaigns, and the use of antibiotics in industrialized countries, the "education" of the immune system has been significantly diminished, and the immune response runs out of control upon stimulation with otherwise innocuous substances.

The data obtained in the non-obese diabetic (NOD) mouse, a laboratory model of T1DM, seems to support the hygiene hypothesis. T1DM can be prevented in NOD mice by infection with bacteria, viruses or parasites [9–12]. It should be noted that T1DM protection does not need these agents to be alive: preparations of dead mycobacteria [13–15], streptococci [16] or parasites [17, 18] can also halt the process that leads to overt T1DM. Thus, microbial components supply the immune information needed to shut off autoimmune diabetes.

3. MECHANISMS OF T1DM PREVENTION

Several mutually non-exclusive mechanisms have been invoked to explain the protection from autoimmune diseases afforded by infections. Microbial epitopes can share sequence homology with regulatory self-epitopes; this "molecular mimicry"

[19] might allow infections (or more specifically, microbial molecules) to activate built-in regulatory networks. Although such a mechanism has been reported to inhibit experimental autoimmune encephalomyelitis [20] and adjuvant arthritis [21], it has not yet been reported for TIDM. In fact, molecular mimicry with microbial antigens seems to accelerate, rather than inhibit TIDM [22]. The contribution of molecular mimicry, and other mechanisms involving the adaptive immune response (antigen competition, bystander suppression or microbial superantigens) to the control of autoimmune diseases by the environment has been recently reviewed elsewhere [1].

4. TOLL-LIKE RECEPTORS

Innate immunity might also play a role in the modulation of TIDM by the environment. Toll-like receptors (TLRs) constitute a family of innate receptors recently identified in mice and humans [23–25]. TLRs were identified based on their homology with the *Drosophila melanogaster* toll receptor, which is involved in the development and the immune response of the fly [26]. Ten different TLRs have been identified thus far in mice and humans [23, 25]. However, none of the TLR knock-out mice described so far has developmental disorders, suggesting that mammalian TLRs lack a role in development or have some degree of redundancy.

Mammalian TLRs are type I transmembrane receptors that share several structural/functional features [23, 25, 27]. They present an extracellular leucine-rich (LRR) repeat whose length varies between different TLRs. LRRs are thought to mediate protein-ligand interactions; they are found in proteins with several functions not restricted to the immune response. The LRR domain of the TLRs is separated from the single transmembrane domain by a characteristic cysteine-rich domain. TLRs also share a cytoplasmic Toll/Interleukin-1 receptor homology (TIR) domain. TIR domains are protein interaction modules shown to recruit adaptor molecules. Finally, all TLRs have at least one MyD88-dependant signaling pathway. MyD88 is a 35 kDa adaptor protein that, through its own C-terminal TIR, interacts with the TIR of activated TLRs [28, 29]. However, MyD88 is not the only adaptor

molecule involved in TLR-triggered signaling. MyD88-independent signaling pathways have also been described [30–32].

Functional studies indicate that TLRs can dimerize, generating both homo and heterodimers [33–35]. Their ability to dimerize and form complexes with other surface molecules like MD2 [36] might explain the structural diversity of TLR ligands. A striking example of ligand diversity is given by TLR4, involved in the recognition of both lipopolysaccharide (LPS) [36] and chlamydial 60 kDa heat shock protein (HSP60) [37]. Although they were initially thought to recognize only pathogen-associated molecules [38], TLRs respond both to self and non-self molecules [39]. Table 1 lists some microbial and host TLR ligands.

5. TLRs AND THE IMMUNE RESPONSE

Dendritic cells (DC) are antigen-presenting cells that trigger and influence several aspects of the immune response, including the differentiation of naïve CD4⁺ T cells into either Th1 or Th2 effector/memory cells [40, 41]. DC express several TLRs whose levels are adjusted according to the state of activation of the cell [42–46], thus by modulating DC activity, TLR ligands could potentially influence the adaptive immune response. TLR activation by self [47–49] or non-self [37, 42, 50–54] TLR ligands promotes DC maturation. Indeed, several experimental reports seem to indicate that the activation of TLRs on DC has strong effects on the immune response.

MyD88 is a pivotal molecule in the signal transduction pathway of TLRs [28, 29]. Schnare and colleagues showed that the immune response triggered by immunization with antigen in CFA, marked by Th1-type cytokines, cytotoxic activity and specific IgG2a in wild type mice, was highly impaired in MyD88-deficient mice [55]. The inability to mount a vigorous Th1 response might have been associated with a defective maturation of DCs in response to the mycobacterial component of CFA, as reported by the authors [55]. Therefore, the triggering of TLR-dependant signaling pathways are needed for the maturation of DC and the induction of a vigorous Th1 response when microbial products are used as adjuvants. Further studies have also included TLRs

Table 1. Self and non-self TLR ligands

TLR	Non-self Ligands	Self Ligands
TLR1	Mycobacterial lipoprotein Triacylated lipopeptides <i>B. burgdoferi</i> OspA	Unknown
TLR2	<i>P. gingivalis</i> LPS Zymosan Peptidoglycan (bacteria) Lipoproteins (bacteria and mycoplasma) <i>T. cruzi</i> GPI anchor <i>B. burgdoferi</i> OspA	HSP60 Surfactant protein A HSP70
TLR3	Poly (I:C) dsRNA	Unknown
TLR4	LPS Respiratory syncytial virus GroEL HSP60 Chlamydia HSP65	HSP60 HSP70 Saturated fatty acids Unsaturated fatty acids Hyaluronic acid Surfactant protein A Fibronectin
TLR5	Flagellin	Unknown
TLR6	Mycoplasma lipoproteins Lipoteichoic acid Peptidoglycan (bacteria)	Unknown
TLR7	Resiquimod Imiquimod	Unknown
TLR8	Resiquimod Imiquimod	Unknown
TLR9	CpG DNA	dsDNA
TLR10	Unknown	Unknown

in the induction of antigen specific Th2 responses [56, 57]. Overall, these results suggest that TLRs are the receptors involved in the adjuvant properties of several microbial preparations [58].

TLR ligands can also be the targets of the adaptive immune response. OspA is an outer-surface lipoprotein from *B. burgdoferi* that activates macrophages through a TLR1 and TLR2, probably complexed in a heterodimer [59]. Vaccination with OspA is being studied as a tool to fight Lyme disease [59]. Strikingly, TLR1 or TLR2-deficient mice do not mount OspA-specific immune responses upon vaccination [59]. Furthermore, humans with reduced TLR1 surface expression on CD4⁺ cells did not mount a detectable immune response to

OspA after repeated vaccination [59]. These results highlight the importance of TLRs for the design of vaccines and demonstrate that TLR ligands can simultaneously work on several components of the immune response, including CD4⁺ T cells.

TLR signaling has been classically associated with the promotion of a Th1, pro-inflammatory, immune response. TLR activation, however, could also lead to the release of Th2 cytokines [56, 57]. Moreover, TLR signaling is needed for the induction of Th2 responses [56, 57, 60, 61] and for the maintenance of B cell memory in humans [62–64].

Thus, microbial antigens activate specific TLRs and in this way influence the immune response directed against them (and the microbe). However,

as we have already mentioned, microbial infection can also lead to the inhibition of autoimmune diabetes; is there a role for TLRs?

6. TLRs AND AUTOIMMUNITY

A healthy immune system harbors self-reactive T and B cells [65–69]. These self-reactive clones are constantly kept under the active control of regulatory cells [70–72]: the removal of the regulators leads to the onset of autoimmune disease [73]. Remarkably, regulatory T cells express several TLRs and are stimulated by TLR ligands like LPS [74]. In addition, upon activation of their own TLRs, DC can have opposite effects on regulatory cell function. DC can secrete immunomodulatory cytokines that favor induction of regulatory cells, like IL-10 [53, 60] or they can inhibit regulatory T-cells via an IL-6-mediated mechanism [75]. Accordingly, several recent papers have shown that TLRs can play a role in the modulation of T1DM by the environment.

7. IMMUNOSTIMULATORY BACTERIAL DNA MOTIFS

Bacterial DNA, like that present in CFA [76], is rich in DNA motifs that stimulate the innate immune system via TLR9-mediated mechanism [77]. These immunostimulatory DNA sequences consist of a central unmethylated CpG dinucleotide flanked by two 5' purines and two 3' pyrimidines [76]; such a sequence is referred to as a CpG motif. We have demonstrated that CpG motifs present in bacterial DNA can inhibit spontaneous diabetes of the NOD mouse [78], but not the more aggressive cyclophosphamide-accelerated diabetes [79]. Prevention of diabetes was characterized by a decrease in insulinitis, and a down-regulation of the spontaneous proliferative T cell responses to HSP60 and to its 437–460 peptide (p277) which characterize NOD diabetes [78]. Moreover, we detected a concomitant increase in IgG2b antibodies to HSP60 and to p277, and not to other islet antigens (GAD or insulin) or to control antigens. The IgG2b isotype of the specific antibodies, together with the decrease in T cell proliferative responses, indicated a shift of the autoimmune process to a Th2-type in treated mice. These results

suggest that immunostimulation by bacterial DNA motifs can modulate spontaneous HSP60 autoimmunity and inhibit NOD diabetes.

8. LPS

LPS from *E. Coli* stimulates innate immunity via TLR4 [23, 25]. Moreover, administration of LPS has been shown to inhibit spontaneous diabetes [80]. Furthermore, LPS-activated B cells express the Fas ligand, secrete TGF β and can induce apoptosis of diabetogenic T cells [81]. The transfer of LPS-activated B cells could temporarily impair APC function and thereby prevented the onset of diabetes in NOD mice, and halted the pathogenic process in an adoptive transfer model of type I diabetes mellitus (T1DM) to NOD/scid mice [81]. However, the transfer of LPS-activated B cells did not promote Th2 responses to β -cell antigens. Thus, B-cell activation through TLR4 by LPS can inhibit spontaneous autoimmune NOD diabetes.

9. DOUBLE STRANDED RNA

Polyriboinosinic:polyribocytidylic acid (poly I:C) is an analogue of viral double-stranded RNA that has been shown to activate the innate immune system via TLR3 [23, 25]. Serreze and co-workers found that the repeated administration of poly I:C, alone or in combination with IL-2, completely prevented the onset of diabetes [82]. However, the therapeutic effect required continuous administration of the immunostimulants since pancreatic insulin content declined and severity of insulinitis increased following cessation of treatment. T cells isolated from Poly I:C-treated mice were capable of suppressing NOD T-cell responses to alloantigens in a mixed lymphocyte culture, indicating that regulatory T cells can be induced in NOD mice by TLR3-mediated signaling pathways.

10. HSP60 AND PEPTIDE P277

Self-HSP60 is an endogenous ligand for TLR2 and TLR4 [83]. The TLR4 molecule does not seem to bind HSP60 directly, but TLR4 is required to

transduce a signal [83–85]. Macrophages exposed to soluble HSP60 secrete pro-inflammatory mediators such as TNF- α , IL-6, IL-12, and nitric oxide [47, 84, 86]. HSP60 can also induce DC maturation and Th1 responses [47, 48]. The pro-inflammatory effects of HSP60, detectable in the blood of pre-diabetic NOD mice at the peak of the autoimmune attack [87], could contribute to the onset of the disease. Thus circulating HSP60 might accelerate β -cell destruction through TLR signaling. However, the same HSP60 molecule has been shown to arrest the diabetogenic process.

HSP60 is a T and B cell antigen in human and NOD autoimmune diabetes [78, 88–90]. Vaccination of NOD mice with HSP60 [79] or its p277 peptide [91] arrested the development of diabetes and even induced remission of overt hyperglycemia [92]. Successful p277 treatment was associated with the down-regulation of spontaneous T-cell reactivity to p277 and with the induction of antibodies to p277 displaying Th2-like isotypes IgG1 and IgG2b [88].

Type I diabetes in humans was also found to be susceptible to immunomodulation by p277 therapy. A double-blind, phase II clinical trial was designed to study the effects of p277 therapy on newly diagnosed patients [93]. The administration of p277 after the onset of clinical diabetes preserved the endogenous levels of C-peptide (which fell in the placebo group) and was associated with lower requirements for exogenous insulin, compatible with the arrest of β -cell destruction. Treatment with p277 was associated with an enhanced Th2 response to HSP60 and p277. Taken together, these results suggest that treatment with HSP60 or its p277 peptide can lead to the induction of HSP60-specific regulators that can control the collective of pathogenic reactivities involved in the progression of autoimmune diabetes.

The apparent contradiction between the pro-inflammatory and the anti-diabetic effects of HSP60 may be resolved by the discovery that HSP60 can directly activate anti-inflammatory effects in T cells by way of an innate receptor. HSP60 and its fragments can regulate the physiology of inflammation itself by acting as ligands for TLR2 in T cells [94]. HSP60 activated human T-cell adhesion to fibronectin, to a degree similar to other activators: IL-2, SDF-1 α and RANTES. T-cell type and state of activation was important; non-activated CD45RA+

and IL-2-activated CD45RO+ T cells responded optimally at low concentrations (0.1–1 ng/ml), but non-activated CD45RO+ T cells required higher concentrations (>1 μ g/ml) of HSP60. T-cell HSP60 signaling was inhibited specifically by a mAb to TLR2, but not by a mAb to TLR4. The human T-cell response to soluble HSP60 depended on PI-3 kinase and PKC signaling, and involved the phosphorylation of Pyk-2. Soluble HSP60 also inhibited actin polymerization and T-cell chemotaxis through ECM-like gels towards the chemokines SDF-1 α or ELC. Exposure to HSP60 could also down-regulate the expression of chemokine receptors CXCR4 and CCR7. Most importantly, HSP60 down-regulated the secretion of IFN γ by activated T cells (unpublished observations). These results suggest that soluble HSP60 (and its fragments), through TLR2-dependent interactions, can down-regulate T-cell behavior and control inflammation. Thus, HSP60 can have both pro- and anti-inflammatory effects on various cell types. HSP60 works as a ligand both for antigen receptors on T cells and B cells (and autoantibodies) and for innate receptors TLR4 and TLR2 on various cells types.

To further examine the contribution of innate immune signaling to autoimmune diabetes, we inserted a TLR4 mutation into NOD mice. As we mentioned above, TLR4 is needed for the activation of macrophages by HSP60 [84, 85]. Mutated TLR4 appears to markedly increase susceptibility to autoimmune type 1 diabetes (Dr. G. Nussbaum, unpublished observations). Apparently TLR4 signaling, whether by endogenous ligands such as HSP60 or foreign ligands such as LPS, can educate the immune system to avoid pathogenic autoimmunity.

11. CONCLUDING REMARKS

TLR-activation has usually been associated with the induction of pro-inflammatory immune responses. Thus the inhibition of T1DM (an inflammatory condition) by TLR-ligands is controversial. Several mechanisms might account for this paradoxical observation. First, the direct activation of regulatory cells via TLR [94]. Second, the activation of TLR-dependant anti-inflammatory responses on effector T cells [94]. Third, by inducing pro-inflamma-

tory responses, TLR ligands might trigger built-in anti-inflammatory responses. Indeed, a controlled inflammatory response has been shown to be necessary for the inhibition of NOD diabetes by CFA [95]. These mechanisms are not exclusive, and further experiments should be directed at determining the contribution of each one of them in the control of diabetes via TLR activation.

Through their interaction with microbial molecules, TLRs sense the environment. The experimental data showing that TLR-mediated activation of the immune system can inhibit the progression of NOD diabetes is in accordance with the hygiene hypothesis. The hygiene hypothesis could then be partially explained based on the "education" of the immune system by microbial TLR ligands. The role played by endogenous TLR ligands is still not clearly understood. However, the preliminary results obtained using NOD mice bearing a non-functional TLR4 might suggest that TLR activation by self ligands is involved in the control of diabetogenic T cells. Maybe the "lesson" thought by exogenous TLR ligands is continuously reinforced via the activation of TLRs by endogenous ligands, like HSP60.

Thus, the stimulation of TLRs with defined TLR ligands might allow us to translate the hygiene hypothesis into immunotherapy. New therapeutic approaches for T1DM aiming at the "re-education" of the immune system might be designed using TLR-ligands, but without the risk of infection with life-threatening pathogens. The initial success of p277 in treating T1DM might then be the first lesson to learn, in a whole new program on the treatment of autoimmune disease.

REFERENCES

1. Bach JF. The effect of infections on susceptibility to autoimmune and allergic diseases. *N Engl J Med* 2002;347:911–20.
2. Hyttinen V, Kaprio J, Kinnunen L, Koskenvuo M, Tuomilehto J. Genetic liability of type 1 diabetes and the onset age among 22,650 young Finnish twin pairs: a nationwide follow-up study. *Diabetes* 2003;52:1052–5.
3. Abbas AK, Lichtman AH, Pober JS. Cellular and Molecular Immunology. Philadelphia: WB Saunders, 1994.
4. Patterson CC, Carson DJ, Hadden DR. Epidemiology of childhood IDDM in Northern Ireland 1989–1994: low incidence in areas with highest population density and most household crowding. Northern Ireland Diabetes Study Group. *Diabetologia* 1996;39:1063–9.
5. Staines A, Bodansky HJ, McKinney PA et al. Small area variation in the incidence of childhood insulin-dependent diabetes mellitus in Yorkshire, UK: links with overcrowding and population density. *Int J Epidemiol* 1997;26:1307–13.
6. Bodansky HJ, Staines A, Stephenson C, Haigh D, Cartwright R. Evidence for an environmental effect in the aetiology of insulin dependent diabetes in a transmigratory population. *BMJ* 1992;304:1020–2.
7. Staines A, Hanif S, Ahmed S, McKinney PA, Shera S, Bodansky HJ. Incidence of insulin dependent diabetes mellitus in Karachi, Pakistan. *Arch Dis Child* 1997;76:121–3.
8. Cohen IR. *Tending Adam's Garden: Evolving the Cognitive Immune Self*. London: Academic, 2000.
9. Cooke A, Tonks P, Jones FM et al. Infection with *Schistosoma mansoni* prevents insulin dependent diabetes mellitus in non-obese diabetic mice. *Parasite Immunol* 1999;21:169–76.
10. Oldstone MB. Viruses as therapeutic agents. I. Treatment of nonobese insulin-dependent diabetes mice with virus prevents insulin-dependent diabetes mellitus while maintaining general immune competence. *J Exp Med* 1990;171:2077–89.
11. Takei I, Asaba Y, Kasatani T et al. Suppression of development of diabetes in NOD mice by lactate dehydrogenase virus infection. *J Autoimmun* 1992;5:665–73.
12. Bras A, Aguas AP. Diabetes-prone NOD mice are resistant to *Mycobacterium avium* and the infection prevents autoimmune disease. *Immunology* 1996;89:20–5.
13. Qin HY, Sadelain MW, Hitchon C, Lauzon J, Singh B. Complete Freund's adjuvant-induced T cells prevent the development and adoptive transfer of diabetes in nonobese diabetic mice. *J Immunol* 1993;150:2072–80.
14. Qin HY, Singh B. BCG vaccination prevents insulin-dependent diabetes mellitus (IDDM) in NOD mice after disease acceleration with cyclophosphamide. *J Autoimmun* 1997;10:271–8.
15. McInerney MF, Pek SB, Thomas DW. Prevention of insulinitis and diabetes onset by treatment with complete Freund's adjuvant in NOD mice. *Diabetes* 1991;40:715–25.
16. Shintani S, Satoh J, Seino H, Goto Y, Toyota T. Mechanism of action of a streptococcal preparation (OK-432) in prevention of autoimmune diabetes in NOD mice. Suppression of generation of effector cells for pancreatic B cell destruction. *J Immunol* 1990;144:136–41.

17. Zaccane P, Fehervari Z, Jones FM et al. Schistosoma mansoni antigens modulate the activity of the innate immune response and prevent onset of type 1 diabetes. *Eur J Immunol* 2003;33:1439–49.
18. Imai S, Tezuka H, Fujita K. A factor of inducing IgE from a filarial parasite prevents insulin-dependent diabetes mellitus in nonobese diabetic mice. *Biochem Biophys Res Commun* 2001;286:1051–8.
19. Abu-Shakra M, Buskila D, Shoenfeld Y. Molecular mimicry between host and pathogen: examples from parasites and implication. *Immunol Lett* 1999;67:147–52.
20. Ruiz PJ, Garren H, Hirschberg DL et al. Microbial epitopes act as altered peptide ligands to prevent experimental autoimmune encephalomyelitis. *J Exp Med* 1999;189:1275–84.
21. Moudgil KD, Kim E, Yun OJ, Chi HH, Brahn E, Sercarz EE. Environmental modulation of autoimmune arthritis involves the spontaneous microbial induction of T cell responses to regulatory determinants within heat shock protein 65. *J Immunol* 2001;166:4237–43.
22. Serreze DV, Ottendorfer EW, Ellis TM, Gauntt CJ, Atkinson MA. Acceleration of type 1 diabetes by a coxsackievirus infection requires a preexisting critical mass of autoreactive T-cells in pancreatic islets. *Diabetes* 2000;49:708–11.
23. Akira S. Mammalian Toll-like receptors. *Curr Opin Immunol* 2003;15:5–11.
24. Imler JL, Hoffmann JA. Toll and Toll-like proteins: an ancient family of receptors signaling infection. *Rev Immunogenet* 2000;2:294–304.
25. Takeda K, Kaisho T, Akira S. Toll-like receptors. *Annu Rev Immunol* 2003;21:335–76.
26. Imler JL, Hoffmann JA. Toll receptors in Drosophila: a family of molecules regulating development and immunity. *Curr Top Microbiol Immunol* 2002;270:63–79.
27. Means TK, Golenbock DT, Fenton MJ. Structure and function of Toll-like receptor proteins. *Life Sci* 2000;68:241–58.
28. Takeuchi O, Akira S. MyD88 as a bottle neck in Toll/IL-1 signaling. *Curr Top Microbiol Immunol* 2002;270:155–67.
29. Janssens S, Beyaert R. A universal role for MyD88 in TLR/IL-1R-mediated signaling. *Trends Biochem Sci* 2002;27:474–82.
30. Yamamoto M, Sato S, Hemmi H et al. TRAM is specifically involved in the Toll-like receptor 4-mediated MyD88-independent signaling pathway. *Nat Immunol* 2003;4:1144–50.
31. Yamamoto M, Sato S, Hemmi H et al. Role of adaptor TRIF in the MyD88-independent toll-like receptor signaling pathway. *Science* 2003;301:640–3.
32. O'Neill LA, Fitzgerald KA, Bowie AG. The Toll-IL-1 receptor adaptor family grows to five members. *Trends Immunol* 2003;24:286–90.
33. Takeuchi O, Sato S, Horiuchi T et al. Cutting edge: role of Toll-like receptor 1 in mediating immune response to microbial lipoproteins. *J Immunol* 2002;169:10–4.
34. Takeuchi O, Kawai T, Muhlratt PF et al. Discrimination of bacterial lipoproteins by Toll-like receptor 6. *Int Immunol* 2001;13:933–40.
35. Ozinsky A, Underhill DM, Fontenot JD et al. The repertoire for pattern recognition of pathogens by the innate immune system is defined by cooperation between toll-like receptors. *Proc Natl Acad Sci USA* 2000;97:13766–71.
36. Akashi S, Saitoh S, Wakabayashi Y et al. Lipopolysaccharide interaction with cell surface Toll-like receptor 4-MD-2: higher affinity than that with MD-2 or CD14. *J Exp Med* 2003;198:1035–42.
37. Costa CP, Kirschning CJ, Busch D et al. Role of chlamydial heat shock protein 60 in the stimulation of innate immune cells by Chlamydia pneumoniae. *Eur J Immunol* 2002;32:2460–70.
38. Medzhitov R. Toll-like receptors and innate immunity. *Nat Rev Immunol* 2001;1:135–45.
39. Beg AA. Endogenous ligands of Toll-like receptors: implications for regulating inflammatory and immune responses. *Trends Immunol* 2002;23:509–12.
40. Boonstra A, Asselin-Paturel C, Gilliet M et al. Flexibility of mouse classical and plasmacytoid-derived dendritic cells in directing T helper type 1 and 2 cell development: dependency on antigen dose and differential toll-like receptor ligation. *J Exp Med* 2003;197:101–9.
41. Mellman I, Steinman RM. Dendritic cells: specialized and regulated antigen processing machines. *Cell* 2001;106:255–8.
42. Krug A, Towarowski A, Britsch S et al. Toll-like receptor expression reveals CpG DNA as a unique microbial stimulus for plasmacytoid dendritic cells which synergizes with CD40 ligand to induce high amounts of IL-12. *Eur J Immunol* 2001;31:3026–37.
43. Liu T, Matsuguchi T, Tsuboi N, Yajima T, Yoshikai Y. Differences in expression of toll-like receptors and their reactivities in dendritic cells in BALB/c and C57BL/6 mice. *Infect Immun* 2002;70:6638–45.
44. Kadowaki N, Ho S, Antonenko S et al. Subsets of human dendritic cell precursors express different toll-like receptors and respond to different microbial antigens. *J Exp Med* 2001;194:863–9.
45. Visintin A, Mazzoni A, Spitzer JH, Wyllie DH, Dower SK, Segal DM. Regulation of Toll-like receptors in human monocytes and dendritic cells. *J Immunol* 2001;166:249–55.
46. Muzio M, Bosisio D, Polentarutti N et al. Differential

- expression and regulation of toll-like receptors (TLR) in human leukocytes: selective expression of TLR3 in dendritic cells. *J Immunol* 2000;164:5998–6004.
47. Flohe SB, Bruggemann J, Lendemans S et al. Human heat shock protein 60 induces maturation of dendritic cells versus a Th1-promoting phenotype. *J Immunol* 2003;170:2340–8.
 48. Bethke K, Staib F, Distler M et al. Different efficiency of heat shock proteins (HSP) to activate human monocytes and dendritic cells: superiority of HSP60. *J Immunol* 2002;169:6141–8.
 49. Termeer C, Benedix F, Sleeman J et al. Oligosaccharides of Hyaluronan activate dendritic cells via toll-like receptor 4. *J Exp Med* 2002;195:99–111.
 50. Tsuji S, Matsumoto M, Takeuchi O et al. Maturation of human dendritic cells by cell wall skeleton of *Mycobacterium bovis* bacillus Calmette-Guerin: involvement of toll-like receptors. *Infect Immun* 2000;68:6883–90.
 51. Hertz CJ, Kiertscher SM, Godowski PJ et al. Microbial lipopeptides stimulate dendritic cell maturation via Toll-like receptor 2. *J Immunol* 2001;166:2444–50.
 52. Michelsen KS, Aicher A, Mohaupt M et al. The role of toll-like receptors (TLRs) in bacteria-induced maturation of murine dendritic cells (DCs). Peptidoglycan and lipoteichoic acid are inducers of DC maturation and require TLR2. *J Biol Chem* 2001;276:25680–6.
 53. Weigt H, Muhlradt PF, Emmendorffer A, Krug N, Braun A. Synthetic mycoplasma-derived lipopeptide MALP-2 induces maturation and function of dendritic cells. *Immunobiology* 2003;207:223–33.
 54. Uehori J, Matsumoto M, Tsuji S et al. Simultaneous blocking of human Toll-like receptors 2 and 4 suppresses myeloid dendritic cell activation induced by *Mycobacterium bovis* bacillus Calmette-Guerin peptidoglycan. *Infect Immun* 2003;71:4238–49.
 55. Schnare M, Barton GM, Holt AC, Takeda K, Akira S, Medzhitov R. Toll-like receptors control activation of adaptive immune responses. *Nat Immunol* 2001;2:947–50.
 56. Eisenbarth SC, Piggott DA, Huleatt JW, Visintin I, Herrick CA, Bottomly K. Lipopolysaccharide-enhanced, toll-like receptor 4-dependent T helper cell type 2 responses to inhaled antigen. *J Exp Med* 2002;196:1645–51.
 57. Dabbagh K, Dahl ME, Stepick-Biek P, Lewis DB. Toll-like receptor 4 is required for optimal development of Th2 immune responses: role of dendritic cells. *J Immunol* 2002;168:4524–30.
 58. Kaisho T, Akira S. Toll-like receptors as adjuvant receptors. *Biochim Biophys Acta* 2002;1589:1–13.
 59. Alexopoulou L, Thomas V, Schnare M et al. Hyporesponsiveness to vaccination with *Borrelia burgdorferi* OspA in humans and in TLR1- and TLR2-deficient mice. *Nat Med* 2002;8:878–84.
 60. Higgins SC, Lavelle EC, McCann C et al. Toll-like receptor 4-mediated innate IL-10 activates antigen-specific regulatory T cells and confers resistance to *Bordetella pertussis* by inhibiting inflammatory pathology. *J Immunol* 2003;171:3119–27.
 61. Kaisho T, Hoshino K, Iwabe T, Takeuchi O, Yasui T, Akira S. Endotoxin can induce MyD88-deficient dendritic cells to support T(h)2 cell differentiation. *Int Immunol* 2002;14:695–700.
 62. Bernasconi NL, Traggiai E, Lanzavecchia A. Maintenance of serological memory by polyclonal activation of human memory B cells. *Science* 2002;298:2199–202.
 63. Bernasconi NL, Onai N, Lanzavecchia A. A role for Toll-like receptors in acquired immunity: up-regulation of TLR9 by BCR triggering in naive B cells and constitutive expression in memory B cells. *Blood* 2003;101:4500–4.
 64. Bourke E, Bosisio D, Golay J, Polentarutti N, Mantovani A. The toll-like receptor repertoire of human B lymphocytes: inducible and selective expression of TLR9 and TLR10 in normal and transformed cells. *Blood* 2003;102:956–63.
 65. Lacroix-Desmazes S, Kaveri SV, Mouthon L et al. Self-reactive antibodies (natural autoantibodies) in healthy individuals. *J Immunol Methods* 1998;216:117–37.
 66. Filion MC, Proulx C, Bradley AJ et al. Presence in peripheral blood of healthy individuals of autoreactive T cells to a membrane antigen present on bone marrow-derived cells. *Blood* 1996;88:2144–50.
 67. Filion MC, Bradley AJ, Devine DV, Decary F, Chartrand P. Autoreactive T cells in healthy individuals show tolerance *in vitro* with characteristics similar to but distinct from clonal anergy. *Eur J Immunol* 1995;25:3123–7.
 68. Martin R, Jaraquemada D, Flerlage M et al. Fine specificity and HLA restriction of myelin basic protein-specific cytotoxic T cell lines from multiple sclerosis patients and healthy individuals. *J Immunol* 1990;145:540–8.
 69. Kellermann SA, McCormick DJ, Freeman SL, Morris JC, Conti-Fine BM. TSH receptor sequences recognized by CD4+ T cells in Graves' disease patients and healthy controls. *J Autoimmun* 1995;8:685–98.
 70. Maloy KJ, Powrie F. Regulatory T cells in the control of immune pathology. *Nat Immunol* 2001;2:816–22.
 71. Tomer Y, Shoenfeld Y. The significance of T suppressor cells in the development of autoimmunity. *J Autoimmun* 1989;2:739–58.
 72. Sakaguchi S. Regulatory T cells: key controllers of immunologic self-tolerance. *Cell* 2000;101:455–8.
 73. Sakaguchi S, Fukuma K, Kuribayashi K, Masuda T. Organ-specific autoimmune diseases induced in mice

- by elimination of T cell subset. I. Evidence for the active participation of T cells in natural self-tolerance; deficit of a T cell subset as a possible cause of autoimmune disease. *J Exp Med* 1985;161:72–87.
74. Caramalho I, Lopes-Carvalho T, Ostler D, Zelenay S, Haury M, Demengeot J. Regulatory T cells selectively express toll-like receptors and are activated by lipopolysaccharide. *J Exp Med* 2003;197:403–411.
 75. Pasare C, Medzhitov R. Toll pathway-dependent blockade of CD4+CD25+ T cell-mediated suppression by dendritic cells. *Science* 2003;299:1033–6.
 76. Krieg AM. CpG motifs in bacterial DNA and their immune effects. *Annu Rev Immunol* 2002;20:709–760.
 77. Hemmi H, Takeuchi O, Kawai T et al. A Toll-like receptor recognizes bacterial DNA. *Nature* 2000;408:740–5.
 78. Quintana FJ, Rotem A, Carmi P, Cohen IR. Vaccination with empty plasmid DNA or CpG oligonucleotide inhibits diabetes in nonobese diabetic mice: modulation of spontaneous 60-kDa heat shock protein autoimmunity. *J Immunol* 2000;165:6148–55.
 79. Quintana FJ, Carmi P, Cohen IR. DNA vaccination with heat shock protein 60 inhibits cyclophosphamide-accelerated diabetes. *J Immunol* 2002;169:6030–5.
 80. Sai P, Rivereau AS. Prevention of diabetes in the nonobese diabetic mouse by oral immunological treatments. Comparative efficiency of human insulin and two bacterial antigens, lipopolysaccharide from *Escherichia coli* and glycoprotein extract from *Klebsiella pneumoniae*. *Diabetes Metab* 1996;22:341–8.
 81. Tian J, Zekzer D, Hanssen L, Lu Y, Olcott A, Kaufman DL. Lipopolysaccharide-activated B cells down-regulate Th1 immunity and prevent autoimmune diabetes in nonobese diabetic mice. *J Immunol* 2001;167:1081–9.
 82. Serreze DV, Hamaguchi K, Leiter EH. Immunostimulation circumvents diabetes in NOD/Lt mice. *J Autoimmun* 1989;2:759–76.
 83. Vabulas RM, Ahmad-Nejad P, da Costa C et al. Endocytosed HSP60s use toll-like receptor 2 (TLR2) and TLR4 to activate the toll/interleukin-1 receptor signaling pathway in innate immune cells. *J Biol Chem* 2001;276:31332–9.
 84. Ohashi K, Burkart V, Flohe S, Kolb H. Cutting edge: heat shock protein 60 is a putative endogenous ligand of the toll-like receptor-4 complex. *J Immunol* 2000;164:558–61.
 85. Habich C, Baumgart K, Kolb H, Burkart V. The receptor for heat shock protein 60 on macrophages is saturable, specific, and distinct from receptors for other heat shock proteins. *J Immunol* 2002;168:569–76.
 86. Kol A, Lichtman AH, Finberg RW, Libby P, Kurt-Jones EA. Cutting edge: heat shock protein (HSP) 60 activates the innate immune response: CD14 is an essential receptor for HSP60 activation of mononuclear cells. *J Immunol* 2000;164:13–7.
 87. Elias D, Markovits D, Reshef T, van der Zee R, Cohen IR. Induction and therapy of autoimmune diabetes in the non-obese diabetic (NOD/Lt) mouse by a 65-kDa heat shock protein. *Proc Natl Acad Sci USA* 1990;87:1576–80.
 88. Elias D, Meilin A, Ablamunits V et al. Hsp60 peptide therapy of NOD mouse diabetes induces a Th2 cytokine burst and downregulates autoimmunity to various beta-cell antigens. *Diabetes* 1997;46:758–64.
 89. Abulafia-Lapid R, Elias D, Raz I, Keren-Zur Y, Atlan H, Cohen IR. T cell proliferative responses of type 1 diabetes patients and healthy individuals to human hsp60 and its peptides. *J Autoimmun* 1999;12:121–9.
 90. Birk OS, Elias D, Weiss AS et al. NOD mouse diabetes: the ubiquitous mouse hsp60 is a beta-cell target antigen of autoimmune T cells. *J Autoimmun* 1996;9:159–66.
 91. Elias D, Reshef T, Birk OS, van der Zee R, Walker MD, Cohen IR. Vaccination against autoimmune mouse diabetes with a T-cell epitope of the human 65-kDa heat shock protein. *Proc Natl Acad Sci USA* 1991;88:3088–91.
 92. Elias D, Cohen IR. Peptide therapy for diabetes in NOD mice. *Lancet* 1994;343:704–6.
 93. Raz I, Elias D, Avron A, Tamir M, Metzger M, Cohen IR. Beta-cell function in new-onset type 1 diabetes and immunomodulation with a heat-shock protein peptide (DiaPep277): a randomised, double-blind, phase II trial. *The Lancet* 2001;358:1749–1753.
 94. Zanin-Zhorov A, Nussbaum G, Franitza S, Cohen IR, Lider O. T cells respond to heat shock protein 60 via TLR2: activation of adhesion and inhibition of chemokine receptors. *Faseb J* 2003;17:1567–9.
 95. Serreze DV, Chapman HD, Post CM, Johnson EA, Suarez-Pinzon WL, Rabinovitch A. Th1 to Th2 cytokine shifts in nonobese diabetic mice: sometimes an outcome, rather than the cause, of diabetes resistance elicited by immunostimulation. *J Immunol* 2001;166:1352–9.