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PRODUCTION OF WHEAT HAPLOIDS USING ANTHER CULTURE AND WHEAT X MAIZE HYBRIDIZATION TECHNIQUES

R. S. Sadasivaiah, B. R. Orshinsky and G. C. Kozub Lethbridge Research Centre, Agriculture and Agri-Food Canada Lethbridge, Alberta, Canada T1J 4B1

Abstract

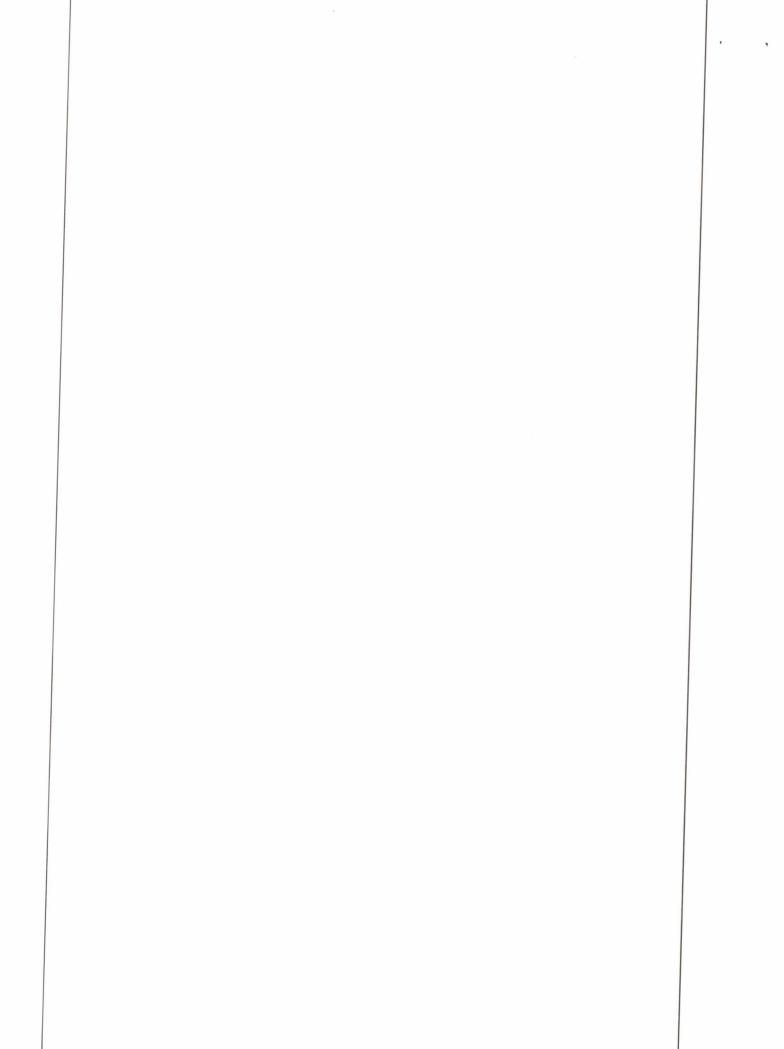
Ten spring wheat breeding lines derived from diverse crosses differed significantly in their response to anther culture and wheat x maize hybridization techniques for the production of haploids. Depending on the genotype, the frequency of embryo induction from cultured anthers ranged from 0.1 to 55.6 per 100 anthers with a mean of 16.3. Plant regeneration from cultured embryos also varied with the genotype. Although the embryos from one line failed to regenerate, those from the nine other lines produced both green and albino plantlets. The frequency of green plants produced per 100 cultured anthers ranged from 0 to 4.73 with a mean of 1.64. Depending on the genotype, 25 to 75% of the anther-derived green plants showed spontaneous chromosome doubling. The wheat x maize hybridization technique was highly effective in producing haploids in all ten breeding lines, although the frequency varied with genotype. The percentage of seed set ranged from 52.7 to 71.6 with a mean of 62.5, but only 19.3% of the seeds contained embryos. About 75% of the cultured embryos developed into green plants. The number of green plants produced per 100 pollinated florets ranged from 2.40 to 12.66 with a mean of 6.29, all having the haploid chromosome number of 21. The line recalcitrant to anther culture produced the highest number of haploids with the wheat x maize hybridization method. There was no evidence of albinism or spontaneous chromosome doubling. The reduced genotype specificity, absence of albinism and ease of application, makes the wheat x maize hybridization technique more efficient than anther culture for the production of haploids in common wheat.

Introduction

Interest in haploids as a genetic and breeding tool is increasing concomitant with the development of more efficient methods for their production. The two major techniques for haploid production in cereals are anther/microspore culture and wide hybridization.

In wheat, use of anther culture in breeding programs is characteristically limited by strong genotype specificity, low frequency of haploids, poor embryo quality, and a high rate of albinism in regenerants (Kasha et al. 1990; Kisana et al., 1993; Orshinsky and Sadasivaiah, 1994, 1997; Lefebvre and Devaux, 1996). Despite these problems, the anther culture technique has been successfully applied in some wheat breeding programs, resulting in new cultivars (Hu et al. 1986; De Buyser et al., 1987; Zhao et al., 1990; R. Graf, 1997, personal comm.).

Production of haploids by wide hybridization followed by the preferential (spontaneous) elimination of the chromosomes of the pollen parent during embryogenesis has been the subject of considerable study since it was first reported from studies of reciprocal interspecific crosses between a wild (*Hordeum bulbosum* L.) and cultivated barley (*Hordeum vulgare* L.) (Symko,



1969: Kasha and Kao, 1970; Lange, 1971; Sadasivaiah and Kasha, 1973; Subrahmanyam and Kasha, 1973). Barclay (1975) recovered a high frequency of haploids of wheat (Triticum aestivum L. var. Chinese Spring) through a similar chromosome elimination process in crosses with H. bulbosum. However, most wheat varieties have very low crossability to H. bulbosum because of the presence of dominant alleles of the Kr1 and Kr2 gene(s) which restrict the application of the *H. bulbosum* technique to haploid production in wheat (Snape et al., 1979; Falk and Kasha, 1981, 1983). The discovery by Laurie and Bennett (1986) that intergeneric crosses between wheat and maize (Zea mays L.) resulted in the production of wheat haploids and that maize appeared insensitive to the action of dominant alleles at wheat crossability loci (Kr1 and Kr2) indicated that wheat x maize crosses could be a valuable alternative to wheat x H bulbosum for production of wheat haploids via chromosome elimination (Laurie and Bennett, 1987, 1988; Suenaga and Nakajima, 1989; Inagaki and Tahir, 1990; Laurie and Reymondie, 1991; Suenaga et al., 1991). Wheat haploids have also been produced by chromosome elimination when pollinated with pollen from such divergent plant species as sorghum (Sorghum bicolor L.), pearl millet (Pennisetum glaucum L.), teosinte (Zea mays L. ssp. mexicana) and eastern gamagrass (Tripsacum dactyloides L.) (Laurie, 1989; Ahmad and Comeau, 1990; Ushiyama et al., 1991; Ohkawa et al., 1992; Riera-Lizarazu and Mujeeb-Kazi, 1993; Inagaki and Bohorova, 1995; Inagaki and Mujeeb-Kazi, 1995; Li et al., 1996).

This paper will report on success rates of wheat haploid production using both anther culture and wheat x maize hybridization techniques and assess their potential as tools in wheat cultivar development programs.

Material and Methods

Plant growth conditions

Ten advanced breeding lines derived from various crosses of soft white spring wheat were used (Table 1). Anther and embryo donor plants were grown in a growth cabinet at 19/16C day/night temperature and 16/8 h photoperiod. Hybrid (CL30/CL34) maize plants used for pollen source were grown in a growth room maintained at 23/20C day/night temperature and 16/8 h photoperiod.

Anther culture

Anthers from primary and secondary florets containing mid-to-late uninucleate microspores were aseptically removed from each of the breeding lines and cultured for 4 to 7 weeks at 30C in the dark in MN6 liquid medium (Orshinsky and Sadasivaiah, 1994). Embryo induction was recorded as the number of embryos produced per 100 anthers. The quality of embryos was visually determined based on size and appearance. Small (less than 1 mm in diameter), white or translucent, and poorly developed embryos were not cultured. Firm, well-developed embryos that were 2-3 mm in diameter were transferred onto MS (Murashige and Skoog, 1962) agar-solidified regeneration medium containing sucrose (20g/l) + 1 mg/l indole-3-acetic acid + 1 mg/l kinetin + 146 mg/l glutamine (Schaeffer et al., 1979), and cultured at 25C with a 16/8h

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photoperiod for 4 to 5 weeks. Regenerated green plantlets were transferred to plastic trays containing Cornell mix (Boodley and Sheldrake, 1977). Once the plantlets were established with good shoot and root systems, they were transferred to root trainers. Plant production was recorded as the number of green plants produced per 100 cultured anthers. The percentages of embryos produced and their subsequent developmental fate (Table 2) were determined for each line and compared statistically (Fleiss, 1981). The CATMOD procedure for categorical modelling in SAS (SAS Institute Inc., 1989) was used to carry out the statistical analyses.

Wheat x maize hybridization procedures

Spikes were hand-emasculated 1-2 days prior to anthesis and covered with glassine bags. One to two days after emasculation the spikes were pollinated with freshly collected maize pollen. On the two consecutive days after pollination the spikes were sprayed with a solution of 75 ppm 2,4-D. On days 3 and 4 after pollination the spikes were sprayed with 300 ppm gibberellic acid (GA3) and re-covered with glassine bags after each spraying.

Embryo rescue

Spikes were harvested at 16-19 days after pollination and the number of intact, non-selfed florets in each was counted and recorded as the number of florets pollinated. Seeds were removed from the florets, sterilized in 70 % ethanol for 30 sec., briefly rinsed in sterile distilled water, then sterilized for 2 min. in 50% Javex with a final 2 min. rinse in sterile distilled water. Embryos were aseptically excised under a stereomicroscope. Small, poorly developed embryos were counted to determine the total embryos formed but were not cultured. Only well-developed embryos were transferred, keeping the embryo axis upwards, to culture vials containing a half-strength MS basal medium supplemented with sucrose (20 g/l), vitamins (myo-inositol, 100 mg/l; thiamine HCl, 1 mg/l; nicotinic acid, 0.5 mg/l; pyridoxine HCl, 0.5 mg/l), amino acids (glutamine, 146 mg/l; glycine, 2 mg/l) and agar (8 g/l). The pH of the medium was adjusted to 5.8 prior to autoclaving.

Vials containing embryos were kept in the dark at 4C for 2 days prior to incubating in the dark at 22C for 1 - 2 weeks and then transferred to a 16/8h light regime at 25C. Germinated and nongerminated embryos were counted, and seedlings with well-developed roots and shoots were transplanted to 4 x 8 root trainers filled with Cornell mix for subsequent growth in a growth cabinet maintained at 15/12 C (day/night) and a 16/8h photoperiod. The percentages of florets with seed, seeds with embryos and embryo developmental fate (Table 3) were determined for each line and compared statistically.

Results

Anther culture

The number of embryos per 100 anthers ranged from 0.1 (B-812) to 55.6 (B-854) with a mean of

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16.3. The two embryos derived from B-812 failed to regenerate, but cultured embryos from the nine other breeding lines produced both green (4.0% to 41.5 %) and albino (21.6% to 35.0%) plantlets. From 36.9 % to 67.7% of the embryos failed to establish plantlets. The frequency of green plantlets produced per 100 cultured anthers from individual lines varied from 0 to 4.73 (Table 2) with an overall mean of 1.64. Depending on the genotype, 25 to 75 % of the anther-derived green plants (mean of 44.1) showed spontaneous chromosome doubling (data not presented). The effect of genotype on the outcomes of culture and regeneration was significant (P<0.001) except for the percentage of cultured embryos with albino shoots (P<0.05).

Wheat x maize hybridization

The percentage of seed set ranged from 52.7 (B-91034) to 71.6 (B-878) with an overall mean of 62.5. However, only 14.7 (B-799) to 28.9% (B-812) of the seed, with a mean of 19.3%, contained an embryo. From 35.2 (B-854) to 81.6% (B-812) of the embryos obtained were considered culturable and 64.4 (B-799) to 86.9% (B-792) of these, with a mean of 75.6%, developed into green plantlets. The frequency of green plantlets per 100 florets ranged from 2.40 (B-854) to 12.66 (B-812) with a mean 6.29. The line recalcitrant to anther culture (B-812) produced the highest number of haploids (12.66) per 100 florets (Table 3). All randomly selected seedlings that were examined cytologically had the haploid chromosome number of 21. There was no evidence of albinism or spontaneous chromosome doubling. The effect of genotype on all the outcomes assessed was significant (P<0.001).

Discussion

Embryo induction frequencies from cultured anthers varied widely among the lines used in the present study. Line B-878 had the highest embryo induction response (55.6 embryos/100 anthers) while B-812 was recalcitrant producing only 0.1embryo/100 anthers. The frequency of plantlet regeneration from cultured embryos also varied with genotype. An overall average of 42.2% (0 to 63.1%) of the cultured embryos regenerated giving a mean yield of 1.64 green (0.47 to 4.73) and 3.24 (0 to 9.73) albino plantlets per 100 anthers. In a study evaluating the anther culture response of 38 different three-way wheat crosses, Kisana et al (1993) also observed significant differences among crosses for plantlet regeneraton (0 to 11.9, mean of 0.6/100 anthers). The strong genotypic effects, low frequency of haploids, and a high rate of albino regenerants limits the use of the anther culture technique for haploid production in wheat breeding programs (De Buyser and Henry, 1986; Henry and De Buyser, 1990; Orshinsky and Sadasivaiah, 1994, 1997; Lefebvre and Devaux, 1996).

All ten of the lines evaluted produced haploids in crosses with maize although they differed significantly in percentage of seed set, embryo formation and plantlet regeneration. The percentage of seed set ranged from 52.7 to 71.6, with a mean of 65.5, but only 19.3% (14.7 to 28.9%) of the seeds contained embryos. From 64.4 to 86.9% of the cultured embryos (mean of 75.6%) regenerated to yield an average of 6.29 (2.40 to 12.66) green plantlets per 100 florets. There was no evidence of albinism or spontaneous chromosome doubling in the regenerants.

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The genotype (B-812) which was recalcitrant to the anther culture method proved to be good parental material for the wheat x maize method. The wheat x maize hybridization technique for wheat haploid production appears to be particularly promising for those genotypes which have proved recalcitrant or respond poorly to anther culture (see also Kisana et al., 1993; Lefebvre and Devaux, 1996; G. Fedak et al., 1997).

Genotypic differences for wheat haploid production efficiency with the maize hybridization method have been reported for both the wheat and maize parents (Suenaga et al., 1991). Laurie and Reymondie (1991), reported that 19 commercial wheat varieties crossed with the maize hybrid 'Seneca 60', produced an average of 20 haploid plants (3 to 22) per 100 pollinated florets. However, Kisana et al (1993) only obtained an average of 4.7 haploid plants (2.7 to 6.3) per 100 florets from wheat x maize crosses. Using 5 maize parents crossed with 18 wheat hybrids, Lefebvre and Devaux (1996) found genotypic differences for haploid production among combinations of parents and the average number of haploids produced per 100 florets was 9.1 which is similar to the frequency (9.5 per 100 florets) reported by Inagaki and Tahir (1990). These results and those obtained in the present study show that the wheat x maize hybridization technique is an effective means of producing wheat haploids from a wide range of genotypes.

In the present study, the wheat x maize method gave a considerably higher yield of haploid plants (6.29/100 pollinated florets) than the anther culture method (1.64/100 cultured anthers). Kisana et al. (1993) and Fedak et al. (1997) also arrived at similar conclusions from their studies of doubled haploid production by both anther culture and wheat x maize hybridization methods. Furthermore, there is potential for further improvement in haploid production frequency with the wheat x maize method by the use of more compatible maize genotypes (Lefebvre and Devaux, 1996) and with the improvement of culture media to rescue poorly developed embryos (Comeau et al., 1992). Another advantage is that wheat x maize-derived plants appear to be cytologically stable whereas some plants derived from anther culture have chromosomal abnormalities (De Buyser and Henry, 1986; Kisana et al., 1993). The reduction or absence of genotype specificity, absence of albinism, greater chromosomal stability of derived plants, reduction in steps and time in culture (Table 4), and the ease with which it can be applied, makes the wheat x maize hybridization technique much more efficient than the anther culture for the production of wheat haploids.

Homozygous doubled haploid lines derived from spontaneous and/or induced chromosome doubling offer the unique advantage of attaining instant homozygosity which enhances the precision of genetic studies and efficiency of selection in breeding programs. The use of doubled haploids as a breeding tool has the potential to greatly improve selection efficiency, reduce research costs, and deliver improved cultivars up to 4 years earlier than would be possible using conventional breeding methods (Hu, et al. 1986; De Buyser et al., 1987). Reduction of the number of generations required for cultivar development is one of the major benefits to be gained by incorporating doubled haploid methods into conventional breeding programs.

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Table 1. Material used in the study

Line	Pedigree
B-353 B-359 B-700x B-788y B-792y B-799	AC Reed//Owens/IDO159 AC Reed//SWS-103/SWS-18 89-B33/AC Reed Dirkwin/Treasure//Blanca/Fielder Dirkwin/Treasure//Blanca/Fielder Dirkwin/8021-V2//Treasure/Blanca
B-812 B-854x B-878 B-91034 Maize hybrid	HY-355/L(Ind)-19//Wadual/L(Ind)-21 89-B33/AC Reed SWS-15/SWS-18//Blanca/Treasure AC Reed/Centennial CL30/CL34

x, y, denotes sister lines

Table 2. Effect of genotype on wheat haploid production with the anther culture technique

Percentage and Standard Error of **Embryos** Cultured Cultured Cultured Embryos/ embryos with embryos with Green plantlets/ that were Line 100 anthers cultured green shoots albino shoots no shoots 100 anthers B-353 10.1 + 0.670.8 + 3.06.6 + 1.925.7 + 3.367.7+3.6 0.47 + 0.14(2341)+(236)(167)(167)(167)(2341)B-359 15.1 + 0.971.5 + 3.011.0 + 2.427.6 + 3.561.4+3.8 1.19 + 0.27(1507)(228)(163)(163)(163)(1507)B-700x 7.9 + 0.671.9 + 3.719.0 + 3.834.3+4.6 46.7 + 4.91.08 + 0.24(1854)(146)(105)(105)(105)(1854)B-788y 26.8+1.2 56.1+2.6 9.6 + 2.135.0 + 3.455.4 + 3.51.45 + 0.33(1311)(351)(197)(197)(197)(1311)B-792y 32.5+1.2 52.2+2.3 4.0+1.230.6 + 2.965.4 + 3.00.67 + 0.21(1487)(483)(252)(252)(252)(1487)B-799 3.5 + 0.474.6+5.3 30.0+6.522.0 + 5.848.0 + 7.10.79 + 0.20(1896)(67)(50)(50)(50)(1896)B-812* 0.1 100.0 0.0 0.0 100.0 0.00 (1615)(2)(2) (2) (2) (1615)B-854x 55.6+1.3 53.9 + 1.713.6+1.6 32.5 + 2.253.9+2.4 4.08 + 0.52(1469)(817)(440)(440)(440)(1469)B-878 14.5 + 0.978.6 + 2.741.5 + 3.721.6 + 3.136.9 + 3.64.73 + 0.54(1542)(224)(176)(176)(176)(1542)B-91034 11.3+0.7 66.8 + 3.025.4+3.4 32.0 + 3.642.6 + 3.81.91 + 0.29(2247)(253)(169)(169)(169)(2247)< 0.001 < 0.001 < 0.001 < 0.05 < 0.001 < 0.001

^{*} B-812 is omitted from statistical analysis when N = 2; x, y, denotes sister lines

⁺ Number in parentheses is the sample size

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Table 3. Effect of genotypes on wheat haploid production with the wheat x maize hybridization technique

Percentage and Standard Error of						
	Florets	Seed v	vith emb.	Total emb	. Cultured emb.with	Green plantlets/
Line	with seed	Total	culturable	cultured	green shoots	100 anthers
B-353	65.5+0.9	19.5+1.0	15.1+0.9	77.3+2.3	80.6+2.5	7.97+0.54
	(2547)+	(1668)	(1668)	(326)	(252)	(2547)
B-359					75.4+3.9	5.14+0.52
						(1791)
B-700x					65.7+4.6	4.60+0.54
					(105)	(1500)
B-788v				69.4+3.8		6.05+0.71
					(100)	(1124)
B-792v					86.9+3.0	
				(168)		
B-799				49.8+3.5		2.85+0.34
						(2354)
B-812					80.1+2.1	12.66+0.69
					(372)	
B-854x				35.2+4.3		2.40+0.41
					(43)	(1376)
B-878				73.1+3.0		
					(155)	(1375)
B-91034					73.6+3.7	
	(2377)			(191)		(2377)
P	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001

x, y, denotes sister lines

⁺ Number in parentheses is the sample size

Table 4. Approximate time required to produce doubled haploids of wheat using the anther culture and wheat x maize hybridization techniques

	Time in	weeks
Activity	Anther culture	Wheat x maize cross
Plant growth prior to anther		
culture or wheat x maize hybridization	7 to 9	8 to 10
Anther culture & embryo induction	4 to 7	N/A
Wheat x maize cross & seed development	N/A	3
Anther-derived embryo regeneration	5	N/A
Wheat x maize-derived embryo regeneration	N/A	4
Seedling establishment & colchicine treatment	7 to 8	5
Plant recovery, growth & seed harvest	12 to 18	12 to 16
Total time required in weeks	35 to 47	32 to 38

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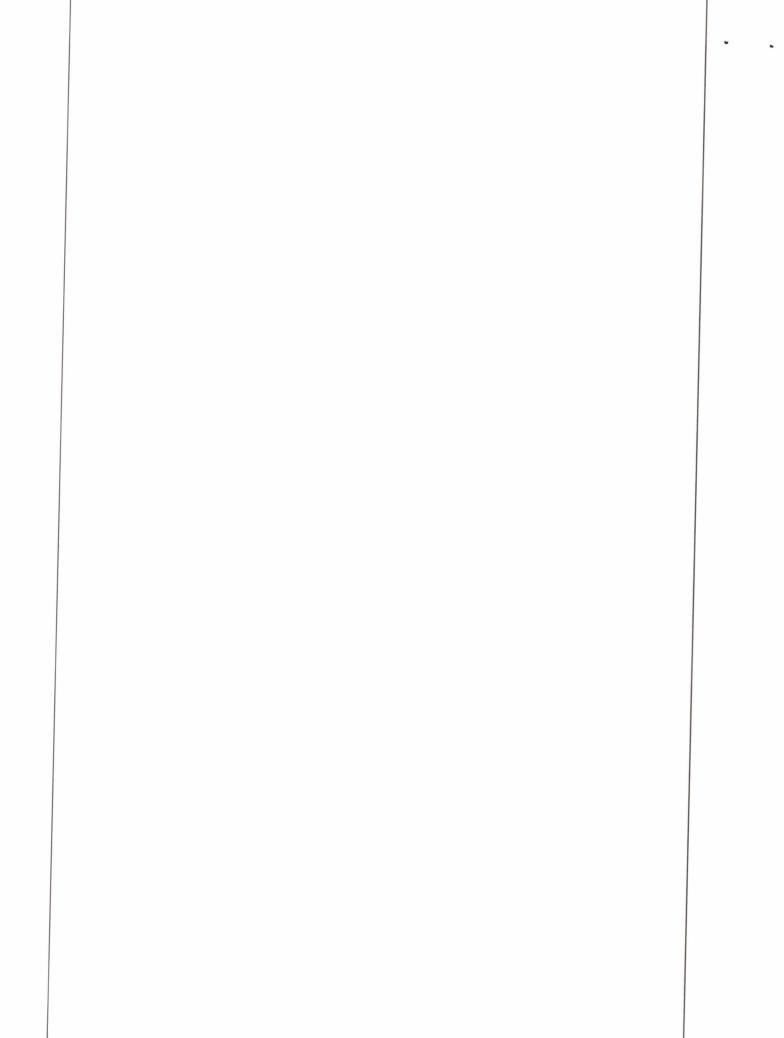
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